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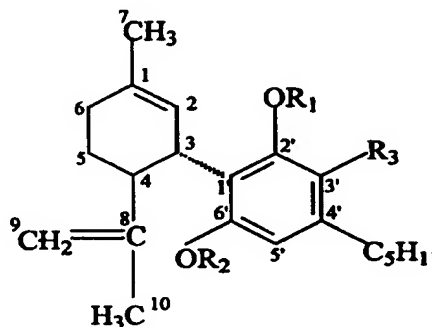
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(54) Title: CANNABINOID AS ANTIOXIDANTS AND NEUROPROTECTANTS

(57) Abstract

Cannabinoids have been found to have antioxidant properties, unrelated to NMDA receptor antagonism. This new found property makes the cannabinoids useful in the treatment and prophylaxis of wide variety of oxidation associated diseases, such as ischemic, age-related, inflammatory and autoimmune diseases. The cannabinoids are found to have particular application as neuroprotectants, for example in limiting neurological damage following ischemic insults, such

as stroke and trauma, or in the treatment of neurodegenerative diseases, such as Alzheimer's disease, Parkinson's disease and HIV dementia. Nonpsychoactive cannabinoids, such as cannabidiol, are particularly advantageous to use because they avoid toxicity that is encountered with psychoactive cannabinoids at high doses useful in the method of the present invention. A particular disclosed class of cannabinoids useful as neuroprotective antioxidants is formula (I) wherein the R group is independently selected from the group consisting of H, CH₃, and COCH₃.



(I)

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CANNABINOIDS AS ANTIOXIDANTS AND NEUROPROTECTANTS

FIELD OF THE INVENTION

The present invention concerns pharmaceutical compounds and compositions that
5 are useful as tissue protectants, such as neuroprotectants and cardioprotectants. The compounds and compositions may be used, for example, in the treatment of acute ischemic neurological insults or chronic neurodegenerative diseases.

BACKGROUND OF THE INVENTION

10 Permanent injury to the central nervous system (CNS) occurs in a variety of medical conditions, and has been the subject of intense scientific scrutiny in recent years. It is known that the brain has high metabolic requirements, and that it can suffer permanent neurologic damage if deprived of sufficient oxygen (hypoxia) for even a few minutes. In the absence of oxygen (anoxia), mitochondrial production of ATP cannot meet the metabolic requirements of the
15 brain, and tissue damage occurs. This process is exacerbated by neuronal release of the neurotransmitter glutamate, which stimulates NMDA (N-methyl-D-aspartate), AMPA (α -amino-3-hydroxy-5-methyl-4-isoxazole propionate) and kainate receptors. Activation of these receptors initiates calcium influx into the neurons, and production of reactive oxygen species, which are potent toxins that damage important cellular structures such as membranes, DNA and enzymes.

20 The brain has many redundant blood supplies, which means that its tissue is seldom completely deprived of oxygen, even during acute ischemic events caused by thromboembolic events or trauma. A combination of the injury of hypoxia with the added insult of glutamate toxicity is therefore believed to be ultimately responsible for cellular death. Hence if the additive insult of glutamate toxicity can be alleviated, neurological damage could also be lessened. Anti-
25 oxidants and anti-inflammatory agents have been proposed to reduce damage, but they often have poor access to structures such as the brain (which are protected by the blood brain barrier).

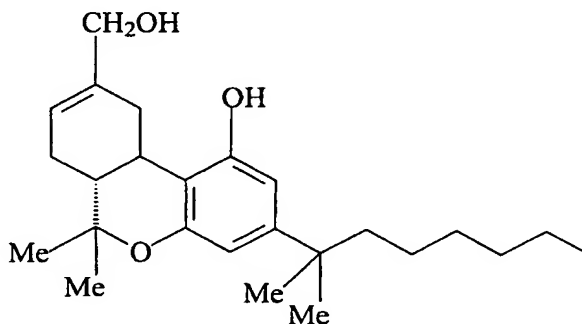
Given the importance of the NMDA, AMPA and kainate receptors in the mechanism of injury, research efforts have focused on using antagonists to these receptors to interfere with the receptor mediated calcium influx that ultimately leads to cellular death and tissue
30 necrosis. *In vitro* studies using cultured neurons have demonstrated that glutamate receptor antagonists reduce neurotoxicity, but NMDA and AMPA/kainate receptor antagonists have different effects. Antagonists to NMDA_A prevent neurotoxicity if present during the glutamate exposure period, but are less effective if added after glutamate is removed. In contrast, AMPA/kainate receptor antagonists are not as effective as NMDA antagonists during the glutamate exposure
35 period, but are more effective following glutamate exposure.

Some of the research on these antagonists has focused on cannabinoids, a subset of which have been found to be NMDA receptor antagonists. U.S. Patent No. 5,538,993 (3S,4S-

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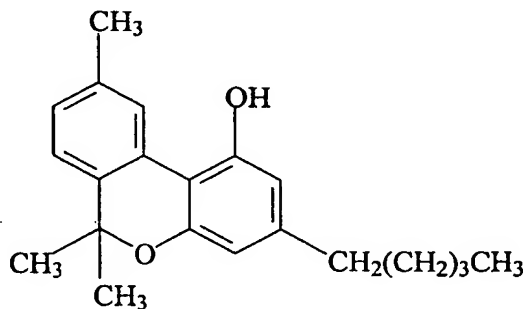
delta-6-tetrahydrocannabinol-7-oic acids), U.S. Patent No. 5,521,215 (stereospecific (+) THC enantiomers), and U.S. Patent No. 5,284,867 (dimethylheptyl benzopyrans) have reported that these cannabinoids are effective NMDA receptor blockers. U.S. Patent No. 5,434,295 discloses that the 1,1 dimethylheptyl (DMH) homolog of [3R,4R]-7-hydroxy- Δ^6 THC (known as HU-210) is a superpotent cannabinoid receptor agonist with cannabinomimetic activity two orders of magnitude greater than the natural Δ^9 THC. The HU-210 dimethylheptyl cannabinoid, has severe side effects, including fatigue, thirst, headache, and hypotension. *J. Pharmacol. Sci.* 60:1433-1457 (1971). Subjects who received this synthetic cannabinoid with a dimethylheptyl group experienced marked psychomotor retardation, and were unwilling or incapable of assuming an erect position.

10 In contrast to HU-210, the (-)(3R,4R) THC-DMH enantiomer (known as HU-211) displays low affinity to the cannabinoid receptors, but retains NMDA receptor antagonist neuroprotective activity.



HU-211

15 THC (tetrahydrocannabinol) is another of the cannabinoids that has been shown to be neuroprotective in cell cultures, but this protection was believed to be mediated by interaction at the cannabinoid receptor, and so would be accompanied by undesired psychotropic side effects.

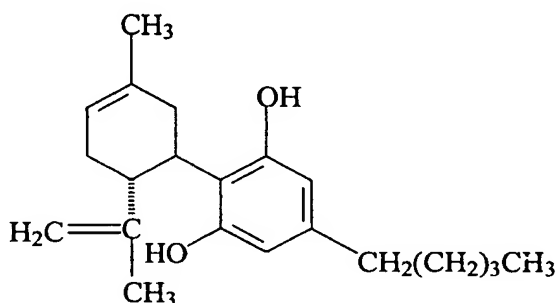


THC

20 Although it has been unclear whether cannabimimetic activity plays a role in neuroprotection against glutamate induced neurological injury, the teaching in this field has clearly

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been that a cannabinoid must at least be an antagonist at the NMDA receptor to have neuroprotective effect. Hence cannabidiol (2-[3-methyl-6-(1-methylethenyl)-2-cyclohexen-1-yl]-5-pentyl-1,3-benzenediol or **CBD**), a cannabinoid devoid of psychoactive effect (*Pharm. Rev.* 38:21-43, 1986), has not been considered useful as a neuroprotectant. Cannabidiol has been studied as an antiepileptic (Carlini et al., *J. Clin. Pharmacol.* 21:417S-427S, 1981; Karler et al., *J. Clin. Pharmacol.* 21:437S-448S, 1981, Consroe et al., *J. Clin. Pharmacol.* 21:428S-436S, 1981), and has been found to lower intraocular pressure (Colasanti et al, *Exp. Eye Res.* 39:251-259, 1984 and *Gen. Pharmac.* 15:479-484, 1984).



10

Cannabidiol (CBD)

No signs of toxicity or serious side effects have been observed following chronic administration of cannabidiol to healthy volunteers (Cunha et al., *Pharmacology* 21:175-185, 1980), even in large acute doses of 700 mg/day (Consroe et al., *Pharmacol. Biochem. Behav.* 40:701-708, 1991) but cannabidiol is inactive at the NMDA receptor. Hence in spite of its potential use in treating glaucoma and seizures, cannabidiol has not been considered a neuroprotective agent that could be used to prevent glutamate induced damage in the central nervous system.

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SUMMARY OF THE INVENTION

It is an object of this invention to provide a new class of antioxidant drugs, that have particular application as neuroprotectants, although they are generally useful in the treatment of many oxidation associated diseases.

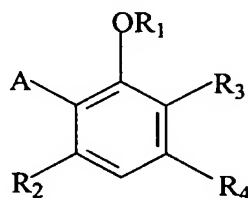
Yet another object of the invention is to provide a subset of such drugs that can be substantially free of psychoactive or psychotoxic effects, are substantially non-toxic even at very high doses, and have good tissue penetration, for example crossing the blood brain barrier.

It has surprisingly been found that cannabidiol and other cannabinoids can function as neuroprotectants, even though they lack NMDA receptor antagonist activity. This discovery was made possible because of the inventor's recognition of a previously unanticipated antioxidant property of the cannabinoids in general (and cannabidiol in particular) that functions completely independently of antagonism at the NMDA, AMPA and kainate receptors. Hence the present

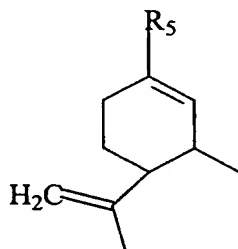
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invention includes methods of preventing or treating diseases caused by oxidative stress, such as neuronal hypoxia, by administering a prophylactic or therapeutically effective amount of a cannabinoid to a subject who has a disease caused by oxidative stress.

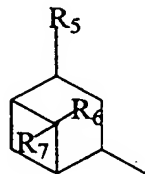
- The cannabinoid may be a cannabinoid other than THC, HU-210, or other potent cannabinoid receptor agonists. The cannabinoid may also be other than HU-211 or any other NMDA receptor antagonist that has previously been reported. A potent cannabinoid receptor agonist is one that has an EC_{50} at the cannabinoid receptor of 50 nM or less, but in more particular embodiments 190 nM or 250 nM or less. In disclosed embodiments the cannabinoid is not psychoactive, and is not psychotoxic even at high doses. In some particularly disclosed
- embodiments, the cannabinoid is selected from the group:



where A is aryl, and particularly



but not a pinene such as:



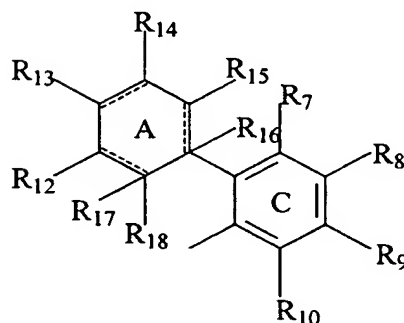
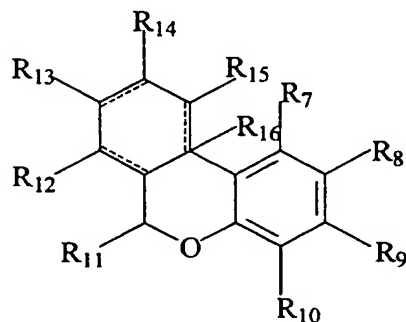
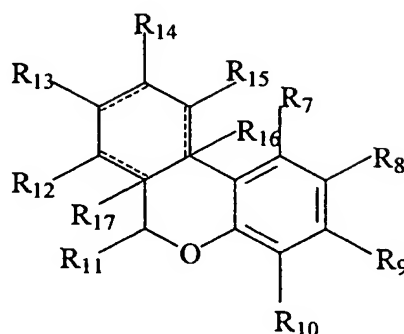
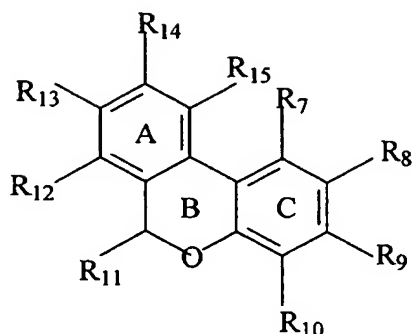
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- and the R_1 - R_5 groups are each independently selected from the groups of hydrogen, lower substituted or unsubstituted alkyl, substituted or unsubstituted carboxyl, substituted or unsubstituted alkoxy, substituted or unsubstituted alcohol, and substituted or unsubstituted ethers, and R_6 - R_7 are H or methyl. In particular embodiments, there are no nitrogens in the rings, and/or no amino
- substitutions in the rings.

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In other embodiments, the cannabinoid is one of the following:



where there can be 0 to 3 double bonds on the A ring, as indicated by the optional double bonds indicated by dashed lines on the A ring. The C ring is aromatic, and the B ring can be a pyran.

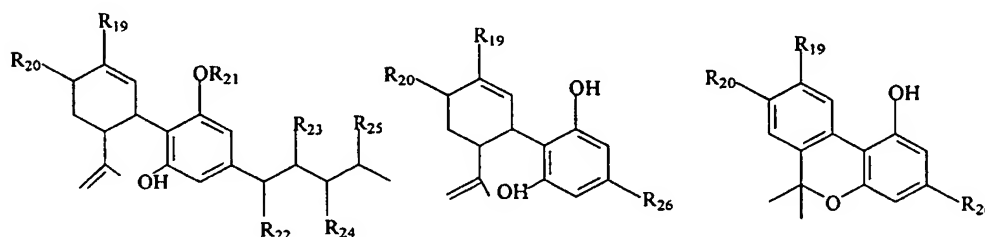
- 5 Particular embodiments are dibenzo pyrans and cyclohexenyl benzenediols. Particular embodiments of the cannabinoids of the present invention may also be highly lipid soluble, and in particular embodiments can be dissolved in an aqueous solution only sparingly (for example 10 mg/ml or less). The octanol/water partition ratio at neutral pH in useful embodiments is 5000 or greater, for example 6000 or greater. This high lipid solubility enhances penetration of the drug
- 10 into the CNS, as reflected by its volume of distribution (V_d) of 1.5 L/kg or more, for example 3.5 L/kg, 7 L/kg, or ideally 10 L/kg or more, for example at least 20 L/kg. Particular embodiments may also be highly water soluble derivatives that are able to penetrate the CNS, for example carboxyl derivatives.

- 15 R_{7-18} are independently selected from the group of H, substituted or unsubstituted alkyl, especially lower alkyl, for example unsubstituted C_1 - C_3 alkyl, hydroxyl, alkoxy, especially lower alkoxy such as methoxy or ethoxy, substituted or unsubstituted alcohol, and unsubstituted or substituted carboxyl, for example COOH or COCH₃. In other embodiments R_{7-18} can also be substituted or unsubstituted amino, and halogen.

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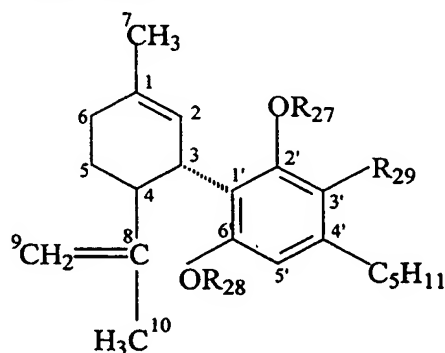
The cannabinoid has substantially no binding to the NMDAr (for example an IC_{50} greater than or equal to $5 \mu M$ or $10 \mu M$), has substantially no psychoactive activity mediated by the cannabinoid receptor (for example an IC_{50} at the cannabinoid receptor of greater than or equal to 300 nM , for example greater than $1 \mu M$ and a K_i greater than 250 nM , especially $500\text{--}1000 \text{ nM}$, for example greater than 1000 nM), and antioxidant activity, as demonstratable by the Fenton reaction or cyclic voltametry.

In other particular embodiments, the cannabinoids are one of the following:



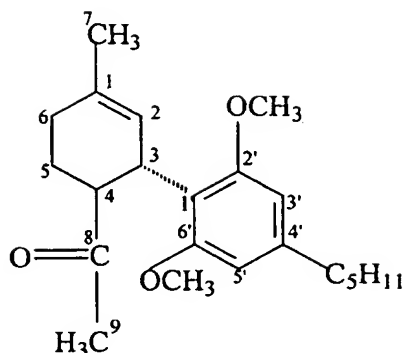
where R_{19} is substituted or unsubstituted alkyl, such as lower alkyl (for example methyl), lower alcohol (such as methyl alcohol) or carboxyl (such as carboxylic acid) and oxygen (as in $=O$); R_{20} is hydrogen or hydroxy; R_{21} is hydrogen, hydroxy, or methoxy; R_{22} is hydrogen or hydroxy; R_{23} is hydrogen or hydroxy; R_{24} is hydrogen or hydroxy; R_{25} is hydrogen or hydroxy; and R_{26} is substituted or unsubstituted alkyl (for example *n*-methyl alkyl), substituted or unsubstituted alcohol, or substituted or unsubstituted carboxy.

In yet other embodiments of the invention, the cannabinoids are



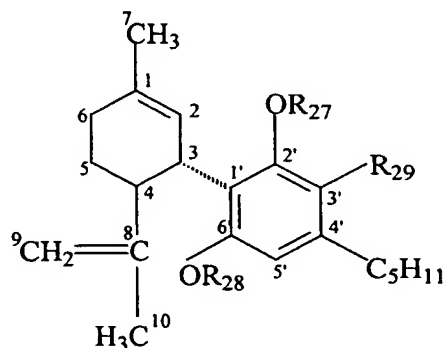
wherein numbering conventions for each of the ring positions are shown, and R_{27} , R_{28} and R_{29} are independently selected from the group consisting of H, unsubstituted lower alkyl such as CH_3 , and carboxyl such as $COCH_3$. Particular examples of nonpsychoactive cannabinoids that fall within this definition are cannabidiol and

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and other structural analogs of cannabidiol.

In more particular embodiments, the cannabinoid is used to prevent or treat an ischemic or neurodegenerative disease in the central nervous system of a subject, by administering to the subject a therapeutically effective amount of a cannabinoid to protect against oxidative injury to the central nervous system. The cannabinoid may be any of the compounds set forth above, or more specifically



wherein R_{27} , R_{28} and R_{29} are independently selected from the group consisting of H, lower alkyl such as CH_3 , and carboxyl such as COCH_3 , and particularly wherein

- a) $R_{27} = R_{28} = R_{29} = \text{H}$
- b) $R_{27} = R_{29} = \text{H}$; $R_{28} = \text{CH}_3$
- c) $R_{27} = R_{28} = \text{CH}_3$; $R_{29} = \text{H}$
- d) $R_{27} = R_{28} = \text{COCH}_3$; $R_{29} = \text{H}$
- e) $R_{27} = \text{H}$; $R_{28} = R_{29} = \text{COCH}_3$

When $R_{27}=R_{28}=R_{29}=\text{H}$, then the compound is cannabidiol. When $R_{27}=R_{29}=\text{H}$ and $R_{28}=\text{CH}_3$, the compound is CBD monomethyl ether. When $R_{27}=R_{28}=\text{CH}_3$ and $R_{29}=\text{H}$, the compound is CBD dimethyl ether. When $R_{27}=R_{28}=\text{COCH}_3$ and $R_{29}=\text{H}$, the compound is CBD diacetate. When $R_{27}=\text{H}$ and $R_{28}=R_{29}=\text{COCH}_3$, the compound is CBD monoacetate. The ischemic or neurodegenerative disease may be, for example, an ischemic infarct, Alzheimer's disease,

The invention also includes identifying and administering antioxidant and neuroprotective compounds (such as cannabidiol) which selectively inhibit the enzyme activity of both 5- and 15-lipoxygenase more than the enzyme activity of 12-lipoxygenase. In addition, such compounds possess low NMDA antagonist activity and low cannabinoid receptor activity. Assays for selecting compounds with the desired effect on lipoxygenase enzymes, and methods for using identified compounds to treat neurological or ischemic diseases are also provided. Such diseases may include a vascular ischemic event in the central nervous system, for example a thromboembolism in the brain, or a vascular ischemic event in the myocardium. Useful administration of the compounds involves administration both during and after an ischemic injury.

25 **BRIEF DESCRIPTION OF THE FIGURES**

FIG. 1B is a graph similar to FIG. 1A, but showing that AMPA/kainate receptor mediated damage (induced by glutamate and the AMPA/kainate receptor potentiating agents cyclothiazide or concanavalin A) is also reduced in a concentration dependent manner by the presence of cannabidiol in the culture medium.

FIG. 2A is a bar graph showing cellular damage (as measured by LDH release) in the presence of glutamate alone (100 μ M Glu), and in the presence of glutamate and 5 μ M cannabidiol (CBD) or 5 μ M THC, and demonstrates that CBD and THC were similarly protective.

FIG. 2B is a bar graph similar to FIG. 2A, but showing the cellular damage assessed in the presence of the cannabinoid receptor antagonist SR 141716A (SR), which was not found to alter the neuroprotective effect of CBD (5 μ M) or THC (5 μ M), indicating the effect is not a typical cannabinoid effect mediated by the cannabinoid receptor.

5 FIG. 3 is a graph showing the reduction oxidation potentials determined by cyclic voltametry for some natural and synthetic cannabinoids, the antioxidant BHT, and the non-cannabinoid anandamide (arachidonyl ethanolamide) which is a ligand for the cannabinoid receptor. The voltage at which initial peaks occur is an indication of antioxidant activity.

10 FIG. 4 is a graph that demonstrates the antioxidant properties of BHT, CBD and THC, by plotting the fluorescence of a fluorescent dye against concentrations of these substances, where declining fluorescence is an indication of greater antioxidant activity.

FIG. 5A is a graph illustrating decreased t-butyl peroxide induced toxicity (as measured by LDH release) in the presence of increasing concentrations of cannabidiol, demonstrating that cannabidiol is an effective antioxidant in living cells.

15 FIG. 5B is a bar graph comparing the antioxidant activity of several antioxidants against glutamate induced toxicity in neurons, showing that CBD has superior antioxidant activity.

FIG. 6A is a graph showing the effect of CBD (as measured by the change in absorbance at 234 nm) on the enzymatic activity of two lipoxygenase enzymes, rabbit 15-LO and porcine 12-LO, which demonstrates that CBD inhibits 15-LO, but not 12-LO enzyme.

20 FIG. 6B is a graph demonstrating that inhibitory effect of CBD on 15-LO is competitive.

FIG. 7A is a graph similar to FIG. 6A, but was performed in whole cells rather than purified enzyme preparations, and shows the effect of CBD (as measured by the change in absorbance at 236 nm) on the enzymatic activity of 5-LO from cultured rat basophilic leukemia cells (RBL-2H3), which demonstrates that CBD inhibits 5-LO.

25 FIG. 7B is a graph showing the effect of CBD (as measured by the change in absorbance at 236 nm) on the formation of 12-HETE (the product of 12-LO) by human leukocytes (12-LO type1).

FIG. 7C is a graph similar to FIG. 7B, showing the effect of CBD (as measured by the change in absorbance at 236 nm) on the formation of 12-HETE by human platelets (12-LO type 2).

FIG. 8 is a bar graph demonstrating that 12-HETE can protect cortical neurons from NMDAr toxicity most effectively when administered during and post ischemia.

35 DETAILED DESCRIPTION OF SOME SPECIFIC EMBODIMENTS

This invention provides antioxidant compounds and compositions, such as pharmaceutical compositions, that include cannabinoids that act as free radical scavengers for use in

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prophylaxis and treatment of disease. The invention also includes methods for using the antioxidants in prevention and treatment of pathological conditions such as ischemia (tissue hypoxia), and in subjects who have been exposed to oxidant inducing agents such as cancer chemotherapy, toxins, radiation, or other sources of oxidative stress. The compositions and methods described herein are also used for preventing oxidative damage in transplanted organs, for inhibiting reoxygenation injury following reperfusion of ischemic tissues (for example in heart disease), and for any other condition that is mediated by oxidative or free radical mechanisms of injury. In particular embodiments of the invention, the compounds and compositions are used in the treatment of ischemic cardiovascular and neurovascular conditions, and neurodegenerative diseases. However the present invention can also be used as an antioxidant treatment in non-neurological diseases.

Molecular oxygen is essential for aerobic organisms, where it participates in many biochemical reactions, including its role as the terminal electron acceptor in oxidative phosphorylation. However excessive concentrations of various forms of reactive oxygen species and other free radicals can have serious adverse biological consequences, including the peroxidation of membrane lipids, hydroxylation of nucleic acid bases, and the oxidation of sulfhydryl groups and other protein moieties. Biological antioxidants include tocopherols and tocotrienols, carotenoids, quinones, bilirubin, ascorbic acid, uric acid, and metal binding proteins. However these endogenous antioxidant systems are often overwhelmed by pathological processes that allow permanent oxidative damage to occur to tissue.

Free radicals are atoms, ions or molecules that contain an unpaired electron, are usually unstable, and exhibit short half-lives. Reactive oxygen species (ROS) is a collective term, designating the oxygen radicals (e.g. O_2^- superoxide radical), which by sequential univalent reduction produces hydrogen peroxide (H_2O_2) and hydroxyl radical (OH^\bullet). The hydroxyl radical sets off chain reactions and can interact with nucleic acids. Other ROS include nitric oxide (NO^\bullet) and peroxy nitrite (NOO^\bullet), and other peroxy (RO_2^\bullet) and alkoxyl (RO^\bullet) radicals. Increased production of these poisonous metabolites in certain pathological conditions is believed to cause cellular damage through the action of the highly reactive molecules on proteins, lipids and DNA. In particular, ROS are believed to accumulate when tissues are subjected to ischemia, particularly when followed by reperfusion.

The pharmaceutical compositions of the present invention have potent antioxidant and/or free radical scavenging properties, that prevent or reduce oxidative damage in biological systems, such as occurs in ischemic/reperfusion injury, or in chronic neurodegenerative diseases such as Alzheimer's disease, HIV dementia, and many other oxidation associated diseases.

35

DEFINITIONS

“Oxidative associated diseases” refers to pathological conditions that result at least in part from the production of or exposure to free radicals, particularly oxyradicals, or reactive oxygen species. It is evident to those of skill in the art that most pathological conditions are multifactorial, and that assigning or identifying the predominant causal factors for any particular condition is frequently difficult. For these reasons, the term “free radical associated disease” encompasses pathological states that are recognized as conditions in which free radicals or ROS contribute to the pathology of the disease, or wherein administration of a free radical inhibitor (e.g. desferroxamine), scavenger (e.g. tocopherol, glutathione) or catalyst (e.g. superoxide dismutase, catalase) is shown to produce detectable benefit by decreasing symptoms, increasing survival, or providing other detectable clinical benefits in treating or preventing the pathological state.

Oxidative associated diseases include, without limitation, free radical associated diseases, such as ischemia, ischemic reperfusion injury, inflammatory diseases, systemic lupus erythematosus, myocardial ischemia or infarction, cerebrovascular accidents (such as a thromboembolic or hemorrhagic stroke) that can lead to ischemia or an infarct in the brain, operative ischemia, traumatic hemorrhage (for example a hypovolemic stroke that can lead to CNS hypoxia or anoxia), spinal cord trauma, Down’s syndrome, Crohn’s disease, autoimmune diseases (e.g. rheumatoid arthritis or diabetes), cataract formation, uveitis, emphysema, gastric ulcers, oxygen toxicity, neoplasia, undesired cellular apoptosis, radiation sickness, and others. The present invention is believed to be particularly beneficial in the treatment of oxidative associated diseases of the CNS, because of the ability of the cannabinoids to cross the blood brain barrier and exert their antioxidant effects in the brain. In particular embodiments, the pharmaceutical composition of the present invention is used for preventing, arresting, or treating neurological damage in Parkinson’s disease, Alzheimer’s disease and HIV dementia; autoimmune neurodegeneration of the type that can occur in encephalitis, and hypoxic or anoxic neuronal damage that can result from apnea, respiratory arrest or cardiac arrest, and anoxia caused by drowning, brain surgery or trauma (such as concussion or spinal cord shock).

As used herein, an “antioxidant” is a substance that, when present in a mixture containing an oxidizable substrate biological molecule, significantly delays or prevents oxidation of the substrate biological molecule. Antioxidants can act by scavenging biologically important reactive free radicals or other reactive oxygen species ($\cdot O_2$, H_2O_2 , $\cdot OH$, $HOCl$, ferryl, peroxy, peroxynitrite, and alkoxy), or by preventing their formation, or by catalytically converting the free radical or other reactive oxygen species to a less reactive species. Relative antioxidant activity can be measured by cyclic voltametry studies of the type disclosed in Example 5 (and FIG. 3), where the voltage (x-axis) is an index of relative antioxidant activity. The voltage at which the first peak occurs is an indication of the voltage at which an electron is donated, which in turn is an index of antioxidant activity.

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“Therapeutically effective antioxidant doses” can be determined by various methods, including generating an empirical dose-response curve, predicting potency and efficacy of a congener by using quantitative structure activity relationships (QSAR) methods or molecular modeling, and other methods used in the pharmaceutical sciences. Since oxidative damage is generally cumulative, there is no minimum threshold level (or dose) with respect to efficacy. However, minimum doses for producing a detectable therapeutic or prophylactic effect for particular disease states can be established.

As used herein, a “cannabinoid” is a chemical compound (such as cannabinol, THC or cannabidiol) that is found in the plant species *Cannabis sativa* (marijuana), and metabolites and synthetic analogues thereof that may or may not have psychoactive properties. Cannabinoids therefore include (without limitation) compounds (such as THC) that have high affinity for the cannabinoid receptor (for example $K_i < 250$ nM), and compounds that do not have significant affinity for the cannabinoid receptor (such as cannabidiol, CBD). Cannabinoids also include compounds that have a characteristic dibenzopyran ring structure (of the type seen in THC) and cannabinoids which do not possess a pyran ring (such as cannabidiol). Hence a partial list of cannabinoids includes THC, CBD, dimethyl heptylpentyl cannabidiol (DMHP-CBD), 6,12-dihydro-6-hydroxy-cannabidiol (described in U.S. Patent No. 5,227,537, incorporated by reference); (3S,4R)-7-hydroxy- Δ^6 -tetrahydrocannabinol homologs and derivatives described in U.S. Patent No. 4,876,276, incorporated by reference; (+)-4-[4-DMH-2,6-diacetoxy-phenyl]-2-carboxy-6,6-dimethylbicyclo[3.1.1]hept-2-en, and other 4-phenylpinene derivatives disclosed in U.S. Patent No. 5,434,295, which is incorporated by reference; and cannabidiol (-)(CBD) analogs such as (-)CBD-monomethylether, (-)CBD dimethyl ether; (-)CBD diacetate; (-)3'-acetyl-CBD monoacetate; and \pm AF11, all of which are disclosed in Consroe et al., *J. Clin. Pharmacol.* 21:428S-436S, 1981, which is also incorporated by reference. Many other cannabinoids are similarly disclosed in Agurell et al., *Pharmacol. Rev.* 38:31-43, 1986, which is also incorporated by reference.

As referred to herein, the term “psychoactivity” means “cannabinoid receptor mediated psychoactivity.” Such effects include, euphoria, lightheadedness, reduced motor coordination, and memory impairment. Psychoactivity is not meant to include non-cannabinoid receptor mediated effects such as the anxiolytic effect of CBD.

The “lipoxygenase enzyme activity” refers to the relative level of lipoxygenase enzyme activity for a particular lipoxgenase, such as 5-, 15- or 12-lipoxygenase, as measured in Example 8. A compound would be said to “selectively inhibit a lipoxgenase enzyme” if the concentration of inhibitor required to reduce enzyme activity by 50% was at least about 5 times less than the amount required to reduce activity of a second lipoxgenase enzyme by the same degree (under the same conditions, i.e. temperature, substrate concentration, etc.)

An “antagonist” is a compound that binds and occupies a receptor without activating it. In the presence of a sufficient concentration of antagonist, an agonist cannot activate

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its receptor. Therefore, antagonists may decrease the neurotoxicity mediated by NMDA (as described in Example 3) or AMPA and Kainate (as described in Example 4).

An "agonist" is a compound that activates a receptor. When the receptor is activated for a longer than normal period of time, this may cause neurotoxicity, as in the case of
5 NMDA, AMPA and kainate receptors (see Examples 3 and 4).

The term "alkyl" refers to a cyclic, branched, or straight chain alkyl group containing only carbon and hydrogen, and unless otherwise mentioned contains one to twelve carbon atoms. This term is further exemplified by groups such as methyl, ethyl, n-propyl, isobutyl, t-butyl, pentyl, pivalyl, heptyl, adamantyl, and cyclopentyl. Alkyl groups can either be
10 unsubstituted or substituted with one or more substituents, e.g. halogen, alkyl, alkoxy, alkylthio, trifluoromethyl, acyloxy, hydroxy, mercapto, carboxy, aryloxy, aryl, arylalkyl, heteroaryl, amino, alkylamino, dialkylamino, morpholino, piperidino, pyrrolidin-1-yl, piperazin-1-yl, or other functionality.

The term "lower alkyl" refers to a cyclic, branched or straight chain monovalent
15 alkyl radical of one to seven carbon atoms. This term is further exemplified by such radicals as methyl, ethyl, n-propyl, i-propyl, n-butyl, t-butyl, i-butyl (or 2-methylpropyl), cyclopropylmethyl, i-amyl, n-amyl, hexyl and heptyl. Lower alkyl groups can also be unsubstituted or substituted, where a specific example of a substituted alkyl is 1,1-dimethyl heptyl.

"Hydroxyl" refers to -OH.

"Alcohol" refers to R-OH, wherein R is alkyl, especially lower alkyl (for example
20 in methyl, ethyl or propyl alcohol). An alcohol may be either linear or branched, such as isopropyl alcohol.

"Carboxyl" refers to the radical -COOH, and substituted carboxyl refers to -COR where R is alkyl, lower alkyl or a carboxylic acid or ester.

The term "aryl" or "Ar" refers to a monovalent unsaturated aromatic carbocyclic
25 group having a single ring (e.g. phenyl) or multiple condensed rings (e.g. naphthyl or anthryl), which can optionally be unsubstituted or substituted with, e.g., halogen, alkyl, alkoxy, alkylthio, trifluoromethyl, acyloxy, hydroxy, mercapto, carboxy, aryloxy, aryl, arylalkyl, heteroaryl, amino, alkylamino, dialkylamino, morpholino, piperidino, pyrrolidin-1-yl, piperazin-1-yl, or other
30 functionality.

The term "alkoxy" refers to a substituted or unsubstituted alkoxy, where an alkoxy has the structure -O-R, where R is substituted or unsubstituted alkyl. In an unsubstituted alkoxy, the R is an unsubstituted alkyl. The term "substituted alkoxy" refers to a group having the structure -O-R, where R is alkyl which is substituted with a non-interfering substituent. The term
35 "arylalkoxy" refers to a group having the structure -O-R-Ar, where R is alkyl and Ar is an aromatic substituent. Arylalkoxys are a subset of substituted alkoxy groups. Examples of useful substituted alkoxy groups are: benzyloxy, naphthyloxy, and chlorobenzyloxy.

The term "aryloxy" refers to a group having the structure -O-Ar, where Ar is an aromatic group. A particular aryloxy group is phenoxy.

The term "heterocycle" refers to a monovalent saturated, unsaturated, or aromatic carbocyclic group having a single ring (e.g. morpholino, pyridyl or furyl) or multiple condensed
5 rings (e.g. indoliziny1 or benzo[b]thienyl) and having at least one heteroatom, defined as N, O, P, or S, within the ring, which can optionally be unsubstituted or substituted with, e.g. halogen, alkyl, alkoxy, alkylthio, trifluoromethyl, acyloxy, hydroxy, mercapto, carboxy, aryloxy, aryl, arylalkyl, heteroaryl, amino, alkylamino, dialkylamino, morpholino, piperidino, pyrrolidin-1-yl, piperazin-1-yl, or other functionality.

10 "Arylalkyl" refers to the groups -R-Ar and -R-HetAr, where Ar is an aryl group. HetAr is a heteroaryl group, and R is a straight-chain or branched chain aliphatic group. Example of arylalkyl groups include benzyl and furfuryl. Arylalkyl groups can optionally be unsubstituted or substituted with, e.g., halogen, alkyl, alkoxy, alkylthio, trifluoromethyl, acyloxy, hydroxy, mercapto, carboxy, aryloxy, aryl, arylalkyl, heteroaryl, amino, alkylamino, dialkylamino,
15 morpholino, piperidino, pyrrolidin-1-yl, piperazin-1-yl, or other functionality.

The term "halo" or "halide" refers to fluoro, bromo, chloro and iodo substituents.

The term "amino" refers to a chemical functionality -NR'R" where R' and R" are independently hydrogen, alkyl, or aryl. The term "quaternary amine" refers to the positively charged group -N⁺R'R", where R'R" and R" are independently selected and are alkyl or aryl. A
20 particular amino group is -NH₂.

A "pharmaceutical agent" or "drug" refers to a chemical compound or composition capable of inducing a desired therapeutic or prophylactic effect when properly administered to a subject.

25 All chemical compounds include both the (+) and (-) stereoisomers, as well as either the (+) or (-) stereoisomer.

Other chemistry terms herein are used according to conventional usage in the art, as exemplified by *The McGraw-Hill Dictionary of Chemical Terms* (1985) and *The Condensed Chemical Dictionary* (1981).

30 The following examples show that both nonpsychoactive cannabidiol, and psychoactive cannabinoids such as THC, can protect neurons from glutamate induced death, by a mechanism independent of cannabinoid receptors. Cannabinoids are also be shown to be potent antioxidants capable of preventing ROS toxicity in neurons.

EXAMPLE 1

35 Preparation of Cannabinoids and Neuronal Cultures

Cannabidiol, THC and reactants other than those specifically listed below were purchased from Sigma Chemical, Co. (St. Louis, MO). Cyclothiazide, glutamatergic ligands and

MK-801 were obtained from Tocris Cookson (UK). Dihydrorhodamine was supplied by Molecular Probes (Eugene, OR). T-butyl hydroperoxide, tetraethylammonium chloride, ferric citrate and sodium dithionite were all purchased from Aldrich (WI). All culture media were Gibco/BRL (MD) products.

5 Solutions of cannabinoids, cyclothiazide and other lipophiles were prepared by evaporating a 10 mM ethanolic solution (under a stream of nitrogen) in a siliconized microcentrifuge tube. Dimethyl sulfoxide (DMSO, less than 0.05% of final volume) was added to ethanol to prevent the lipophile completely drying onto the tube wall. After evaporation, 1 ml of culture media was added and the drug was dispersed using a high power sonic probe. Special
10 attention was used to ensure the solution did not overheat or generate foam. Following dispersal, all solutions were made up to their final volume in siliconized glass tubes by mixing with an appropriate quantity of culture media.

 Primary neuronal cultures were prepared according to the method of Ventra et al. (J. Neurochem. 66:1752-1761, 1996). Fetuses were extracted by Cesarian section from a 17 day
15 pregnant Wistar rat, and the fetal brains were placed into phosphate buffered saline. The cortices were then dissected out, cut into small pieces and incubated with papain for nine minutes at 37°C. After this time the tissue was dissociated by passage through a fire polished Pasteur pipette, and the resultant cell suspension separated by centrifugation over a gradient consisting of 10 mg/ml bovine serum albumin and 10 mg/ml ovomucoid (a trypsin inhibitor) in Earls buffered salt solution. The
20 pellet was then re-suspended in high glucose, phenol red free Dulbecco's modified Eagles medium containing 10% fetal bovine serum, 2 mM glutamine, 100 IU penicillin, and 100 µg/ml streptomycin (DMEM). Cells were counted, tested for vitality using the trypan blue exclusion test and seeded onto poly-D-lysine coated 24 multiwell plates. After 96 hours, 10 µM fluoro-deoxyuridine and 10 µM uridine were added to block glial cell growth. This protocol resulted in a
25 highly neuron-enriched culture.

EXAMPLE 2

Preparation of Astrocytes and Conditioned Media

 Astrocyte conditioned DMEM was used throughout the AMPA/kainate toxicity
30 procedure and following glutamate exposure in the NMDAr mediated toxicity protocol. Media was conditioned by 24 hour treatment over a confluent layer of type I astrocytes, prepared from two day old Wistar rat pups. Cortices were dissected, cut into small pieces, and enzymatically digested with 0.25% trypsin. Tissue was then dissociated by passage through a fire polished Pasteur pipette and the cell suspension plated into untreated 75 cm² T-flasks. After 24 hours the media was
35 replaced and unattached cells removed. Once astrocytes achieved confluence, cells were divided into four flasks. Media for experiments was conditioned by a 24 hour exposure to these astrocytes,

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after which time it was frozen at -20°C until use. Astrocyte cultures were used to condition DMEM for no longer than two months.

EXAMPLE 3

NMDA Mediated Toxicity Studies

5 Glutamate neurotoxicity can be mediated by NMDA, AMPA or kainate receptors. To examine NMDA mediated toxicity, cultured neurons (cultured for 14-18 days) were exposed to 250 μ M glutamate for 10 minutes in a magnesium free saline solution. The saline was composed of 125 mM NaCl, 25 mM glucose, 10 mM HEPES (pH 7.4), 5 mM KCl, 1.8 mM calcium chloride
10 and 5% bovine serum albumin. Following exposure, cells were washed twice with saline, and incubated for 18 hours in conditioned DMEM. The level of lactate dehydrogenase (LDH) in the media was used as an index of cell injury.

 Toxicity was completely prevented by addition of the NMDA antagonist, MK-801 (500 nM, data not shown). However, FIG. 1A shows that cannabidiol also prevented neurotoxicity
15 (maximum protection $88 \pm 9\%$) with an EC_{50} of 2-4 μ M (specifically about 3.5 μ M).

EXAMPLE 4

AMPA and Kainate Receptor Mediated Toxicity Studies

 Unlike NMDA receptors, which are regulated by magnesium ions, AMPA/kainate
20 receptors rapidly desensitize following ligand binding. To examine AMPA and kainate receptor mediated toxicity, neurons were cultured for 7-13 days, then exposed to 100 μ M glutamate and 50 μ M cyclothiazide (used to prevent AMPA receptor desensitization). Cells were incubated with glutamate in the presence of 500 nM MK-801 (an NMDA antagonist) for 18-20 hours prior to analysis. Specific AMPA and kainate receptor ligands were also used to separately examine the
25 effects of cannabinoids on AMPA and kainate receptor mediated events. Fluorowillardiine (1.5 μ M) was the AMPA agonist and 4-methyl glutamate (10 μ M) was the kainate agonist used to investigate receptor mediated toxicity. When specifically examining kainate receptor activity, cyclothiazide was replaced with 0.15 mg/ml Concanavalin-A.

 Cannabidiol protection against AMPA/kainate mediated neurotoxicity is illustrated
30 in FIG. 1B, where LDH in the media was used as an index of cell injury. The neuroprotective effect of cannabidiol was similar to that observed in the NMDA mediated toxicity model (FIG. 1A). Cannabidiol prevented neurotoxicity (maximum protection $80 \pm 17\%$) with an EC_{50} of 2-4 μ M (specifically about 3.3 μ M). Comparable results were obtained with either the AMPA receptor ligand, fluorowillardiine or the kainate receptor specific ligand, 4-methyl-glutamate (data not
35 shown). Hence cannabidiol protects similarly against toxicity mediated by NMDA, AMPA or kainate receptors.

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Unlike cannabidiol, THC is a ligand (and agonist) for the brain cannabinoid receptor. The action of THC at the cannabinoid receptor has been proposed to explain the ability of THC to protect neurons from NMDAr toxicity *in vitro*. However in AMPA/kainate receptor toxicity assays, THC and cannabidiol were similarly protective (FIG. 2A), indicating that
5 cannabinoid neuroprotection is independent of cannabinoid receptor activation. This was confirmed by inclusion of cannabinoid receptor antagonist SR-141716A in the culture media (SR in FIG. 2B). See Mansbach et al., *Psychopharmacology* 124:315-22, 1996, for a description of SR-141716A. Neither THC nor cannabidiol neuroprotection was affected by cannabinoid receptor antagonist (FIG. 2B).

10

EXAMPLE 5

Cyclic Voltametry Studies of ReDox Potentials

To investigate whether cannabinoids protect neurons against glutamate damage by reacting with ROS, the antioxidant properties of cannabidiol and other cannabinoids were assessed.

15 Cyclic voltametry, a procedure that measures the ability of a compound to accept or donate electrons under a variable voltage potential, was used to measure the oxidation potentials of several natural and synthetic cannabinoids. These studies were performed with an EG&G Princeton Applied Research potentiostat/galvanostat (Model 273/ PAR 270 software, NJ). The working electrode was a glassy carbon disk with a platinum counter electrode and silver/silver chloride
20 reference. Tetraethylammonium chloride in acetonitrile (0.1 M) was used as an electrolyte. Cyclic voltametry scans were done from + 0 to 1.8 V at scan rate of 100 mV per second. The reducing ability of cannabidiol (CBD), THC, HU-211, and BHT were measured in this fashion. Anandamide, a cannabinoid receptor ligand without a cannabinoid like structure, was used as a non-responsive control. Each experiment was repeated twice with essentially the same results.

25 Cannabidiol, THC and the synthetic cannabinoid HU-211 all donated electrons at a similar potential as the antioxidant BHT. Anandamide (arachidonyl ethanolamide) did not undergo oxidation at these potentials (FIG. 3). Several other natural and synthetic cannabinoids, including cannabidiol, nabilone, and levantantradol were also tested, and they too exhibited oxidation profiles similar to cannabidiol and THC (data not shown).

30

EXAMPLE 6

Iron Catalyzed Dihydrorhodamine Oxidation (Fenton Reaction)

The ability of cannabinoids to be readily oxidized, as illustrated in Example 5, indicated they possess antioxidant properties comparable to BHT. The antioxidant activity of BHT
35 was examined in a Fenton reaction, in which iron is catalyzed to produce ROS. Cannabidiol (CBD) and tetrahydrocannabinol (THC) were evaluated for their ability to prevent oxidation of dihydrorhodamine to the fluorescent compound rhodamine. Oxidant was generated by ferrous

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catalysis (dithionite reduced ferric citrate) of t-butyl hydroperoxide in a 50:50 water:acetonitrile (v/v) solution. Dihydrorhodamine (50 μ M) was incubated with 300 μ M t-butyl hydroperoxide and 0.5 μ M iron for 5 minutes. After this time, oxidation was assessed by spectrofluorimetry (Excit=500 nm, Emiss=570 nm). Various concentrations of cannabinoids and BHT were included
5 to examine their ability to prevent dihydrorhodamine oxidation.

Cannabidiol, THC and BHT all prevented dihydrorhodamine oxidation in a similar, concentration dependent manner (FIG. 4), indicating that cannabinoids have antioxidant potency comparable to BHT.

To confirm that cannabinoids act as antioxidants in the intact cell, neurons were
10 also incubated with the oxidant t-butyl hydroperoxide and varying concentrations of cannabidiol (FIG. 5A). The t-butyl hydroperoxide oxidant was chosen for its solubility in both aqueous and organic solvents, which facilitates oxidation in both cytosolic and membrane cell compartments. Cell toxicity was assessed 18-20 hours after insult by measuring lactate dehydrogenase (LDH) release into the culture media. All experiments were conducted with triple or quadruple values at
15 each point and all plates contained positive (glutamate alone) and baseline controls. The assay was validated by comparison with an XTT based metabolic activity assay. As shown in FIG. 5A, cannabidiol protected neurons against ROS toxicity in a dose related manner, with an EC_{50} of about 6 μ M. The maximum protection observed was $88 \pm 9\%$.

Cannabidiol was also compared with known antioxidants in an AMPA/kainate
20 toxicity protocol. Neurons were exposed to 100 μ M glutamate and equimolar (5 μ M) cannabidiol, α -tocopherol, BHT or ascorbate (FIG. 5B). Although all of the antioxidants attenuated glutamate toxicity, cannabidiol was significantly more protective than either α -tocopherol or ascorbate. The similar antioxidant abilities of cannabidiol and BHT in this chemical system (FIG. 4), and their comparable protection in neuronal cultures (FIG. 5B), implies that cannabidiol neuroprotection is
25 due to an antioxidant effect.

EXAMPLE 7

In vivo Rat Studies

The middle cerebral artery of chloral hydrate anesthetized rats was occluded by
30 insertion of suture thread into it. The animals were allowed to recover from the anesthetic and move freely for a period of two hours. After this time the suture was removed under mild anesthetic and the animals allowed to recover for 48 hours. Then the animals were tested for neurological deficits, sacrificed, and the infarct volume calculated. To examine the infarct volume, animals were anesthetized, ex-sanguinated, and a metabolically active dye (3-phenyl tetrazolium
35 chloride) was pumped throughout the body. All living tissues were stained pink by the dye, while morbid regions of infarcted tissue remained white. Brains were then fixed for 24 hours in formaldehyde, sliced and the infarct volumes measured.

One hour prior to induction of ischemia 20 mg/kg of cannabidiol was administered by intra-peritoneal injection (ip) in a 90% saline : 5% emulphor 620 (emulsifier) : 5% ethanol vehicle. A second ip 10 mg/kg dose of cannabidiol was administered 8 hours later using the same vehicle. Control animals received injections of vehicle without drug. IV doses would be expected to be 3-5 times less because of reduction of first pass metabolism.

The infarct size and neurological assessment of the test animals is shown Table 1.

Table 1: Cannabidiol protects rat brains from ischemia damage

| Animal | Volume of Infarct (mm3) | | Behavioral Deficit Score | |
|--------|-------------------------|---------|--------------------------|---------|
| | Drug | Control | Drug | Control |
| 1 | 108.2 | 110.5 | 3 | 2 |
| 2 | 83.85 | 119.6 | 4 | 4 |
| 3 | 8.41 | 118.9 | 3 | 4 |
| 4 | 75.5 | 177.7 | 1 | 4 |
| 5 | 60.53 | 33.89 | 1 | 3 |
| 6 | 27.52 | 255.5 | 1 | 5 |
| 7 | 23.16 | 143 | 1 | 4 |
| Mean | 55.3 | 137.0 | 2.0 | 3.7 |
| SEM | 13.8 | 25.7 | 0.5 | 0.4 |
| | p=0.016 significant | | p=0.015 significant | |

*Neurological scoring is performed on a subjective 1-5 scale of impairment. 0 = no impairment, 5 = severe (paralysis)

10

This data shows that infarct size was approximately halved in the animals treated with cannabidiol, which was also accompanied by a substantial improvement in the neurological status of the animal.

These studies with the nonpsychotropic marijuana constituent, cannabidiol, demonstrate that protection can be achieved against both glutamate neurotoxicity and free radical induced cell death. THC, the psychoactive principle of cannabis, also blocked glutamate neurotoxicity with a potency similar to cannabidiol. In both cases, neuroprotection is unaffected by the presence of a cannabinoid receptor antagonist. These results therefore surprisingly demonstrate that cannabinoids can have useful therapeutic effects that are not mediated by cannabinoid receptors, and therefore are not necessarily accompanied by psychoactive side effects. Cannabidiol also acts as an anti-epileptic and anxiolytic, which makes it particularly useful in the treatment of neurological diseases in which neuroanatomic defects can predispose to seizures (e.g. subarachnoid hemorrhage).

A particular advantage of the cannabinoid compounds of the present invention is that they are highly lipophilic, and have good penetration into the central nervous system. The volume of distribution of some of these compounds is at least 100 L in a 70 kg person (1.4 L/kg),

25

more particularly at least 250 L, and most particularly 500 L or even 700 L in a 70 kg person (10 L/kg). The lipophilicity of particular compounds is also about as great as that of THC, cannabidiol or other compounds that have excellent penetration into the brain and other portions of the CNS.

Cannabinoids that lack psychoactivity or psychotoxicity are particularly useful
5 embodiments of the present invention, because the absence of such side effects allows very high
doses of the drug to be used without encountering unpleasant side effects (such as dysphoria) or
dangerous complications (such as obtundation in a patient who may already have an altered mental
status). For example, therapeutic antioxidant blood levels of cannabidiol can be 5-20 mg/kg,
without significant toxicity, while blood levels of psychoactive cannabinoids at this level would
10 produce obtundation, headache, conjunctival irritation, and other problems. Particular examples of
the compounds of the present invention have low affinity to the cannabinoid receptor, for example a
 K_i of greater than 250 nM, for example $K_i \geq 500$ -1000 nM. A compound with a $K_i \geq 1000$ nM is
particularly useful, which compound has essentially no psychoactivity mediated by the cannabinoid
receptor.

15 Cannabidiol blocks glutamate toxicity with equal potency regardless of whether the
insult is mediated by NMDA, AMPA or kainate receptors. Cannabidiol and THC have been shown
to be comparable to the antioxidant BHT, both in their ability to prevent dihydropyridine
oxidation and in their cyclic voltametric profiles. Several synthetic cannabinoids also exhibited
profiles similar to the BHT, although anandamide, which is not structurally related to cannabinoids,
20 did not. These findings indicate that cannabinoids act as antioxidants in a non-biological situation,
which was confirmed in living cells by showing that cannabidiol attenuates hydroperoxide induced
neurotoxicity. The potency of cannabidiol as an antioxidant was examined by comparing it on an
equimolar basis with three other commonly used compounds.

In the AMPA/kainate receptor dependent neurotoxicity model, cannabidiol
25 neuroprotection was comparable to the potent antioxidant, BHT, but significantly greater than that
observed with either α -tocopherol or ascorbate. This unexpected superior antioxidant activity (in
the absence of BHT tumor promoting activity) shows for the first time that cannabidiol, and other
cannabinoids, can be used as antioxidant drugs in the treatment (including prophylaxis) of oxidation
associated diseases, and is particularly useful as a neuroprotectant. The therapeutic potential of
30 nonpsychoactive cannabinoids is particularly promising, because of the absence of psychotoxicity,
and the ability to administer higher doses than with psychotropic cannabinoids, such as THC.
Previous studies have also indicated that cannabidiol is not toxic, even when chronically
administered to humans or given in large acute doses (700 mg/day).

EXAMPLE 8

Effect of Cannabidiol on Lipxygenase Enzymes

This example describes *in vitro* and *in vivo* assays to examine the effect of cannabidiol (CBD) on three lipxygenase (LO) enzymes: 5-LO, 12-LO and 15-LO.

5

In vitro Enzyme assay

The ability of CBD to inhibit lipxygenase was examined by measuring the time dependent change in absorption at 234 nm following addition of 5U of each lipxygenase (rabbit 15-LO purchased from Biomol (PA), porcine 12-LO purchased from Cayman chemicals (MI)) to a solution containing 10 μ M (final concentration) linoleic acid.

Enzyme studies were performed using a u.v. spectrophotometer and a 3 ml quartz cuvette containing 2.5 ml of a stirred solution of 12.5 μ M sodium linoleic acid (sodium salt) in solution A (25 mM Tris (pH 8.1), 1 mM EDTA 0.1% methyl cellulose). The reaction was initiated by addition of 0.5 ml enzyme solution (10 U/ml enzyme in solution A) and recorded for 60 seconds. Lipxygenase exhibits non-Michaelis-Menten kinetics, an initial "lag" (priming) phase followed by a linear phase which is terminated by product inhibition. These complications were reduced by assessing enzyme activity (change in absorption) over the "steepest" 20 second period in a 60 second run time. Recordings examined the absorption at 234 nm minus the value at a reference wavelength of 280 nm. Linoleic acid was used as the substrate rather than arachidonic acid, because the products are less inhibitory to the enzyme, thereby providing a longer "linear phase".

Cell purification and separation

Human platelets and leukocytes were purified from buffy coat preparations (NIH Blood Bank) using a standard Ficoll based centrifugation method used in blood banks. Prior to use, cells were washed three times to eliminate contaminating cell types. Cultured rat basophilic leukemia cells (RBL-2H3) were used as a source of 5-lipxygenase.

In vivo Determination of Lipxygenase activity

Cells were incubated with arachidonic acid and stimulated with the calcium ionophore A23187. Lipids were extracted and separated by reverse phase HPLC. Product formation was assessed as the area of a peak that co-eluted with an authentic standard, had a greater absorbance at 236 nm than at either 210 or 280 nm, and the formation of which was inhibited by a lipxygenase inhibitor.

Cell pellets were triturated in DMEM culture media, aliquoted and pre-incubated for 15 minutes with 20 μ M arachidonic acid and varying concentrations of cannabidiol and/or 40 μ M nordihydroguaiaretic acid (a lipxygenase inhibitor). Platelets and leukocytes were also pre-

incubated with 80 μ M manoalide (Biomol) to prevent phospholipase A2 activation. Product formation was initiated by addition of 5 μ M A23187 and incubation for 10 minutes at 37°C. At the end of the incubation, the reaction was stopped by addition of 15% 1M HCl and 10 ng/ml prostaglandin B2 (internal standard). Lipids were extracted with 1 volume of ethyl ether, which
5 was dried under a stream of nitrogen. Samples were reconstituted in 50% acetonitrile:50% H₂O and separated by reverse phase HPLC using a gradient running from 63% acetonitrile: 37% H₂O:0.2% acetic acid to 90% acetonitrile (0.2% acetic acid) over 13 minutes.

Measurement of NMDAr toxicity

10 The ability of 12-HETE (12-(s)-hydroxy-eicosatetraenoic acid, the product of the action of 12-lipoxygenase on arachidonic (eicosatetraenoic) acid) to protect cortical neurons from NMDAr toxicity was measured as described in Example 3. The 12-HETE (0.5 μ g/ml) was added either during ischemia (co-incubated with the glutamate), during post-ischemia (co-incubated with the DMEM after washing the cells), or during both ischemia and post-ischemia.

15

Results

Using semi-purified enzyme preparations, the effect of CBD on rabbit 15-LO and porcine 12-LO was compared. As shown in FIGS. 6A and B, CBD is a potent competitive inhibitor of 15-LO with an EC₅₀ of 598 nM. However, CBD had no effect on the 12-LO enzyme.

20

Using whole cell preparations, the effect of CBD on 5- and 12-LO enzymes was investigated. As shown in FIG. 7A, CBD inhibited 5-LO in cultured rat basophilic leukemia cells (RBL-2H3) with an EC₅₀ of 1.92 μ M. However, CBD had no effect on 12-LO, as monitored by the production of 12-HETE (the product of 12-LO), in either human leukocytes or platelets (FIGS. 7B and C). The leukocyte 12-LO is similar, while the platelet 12-LO is structurally and
25 functionally different, from the porcine 12-LO used in the *in vitro* enzyme study.

The ability of 12-HETE to protect cortical neurons from NMDAr toxicity is shown in FIG. 8. To achieve best protection from NMDAr toxicity, 12-HETE was administered both during and post ischemia.

Therefore, CBD serves as a selective inhibitor of at least two lipoxygenase
30 enzymes, 5-LO and 15-LO, but had no effect on 12-LO. Importantly, this is the first demonstration (FIG. 8) that the 12-LO product 12-HETE can play a significant role in protecting neurons from NMDAr mediated toxicity. Although the mechanism of this protection is unknown at the present time, 12-HETE is known to be an important neuromodulator, due to its ability to influence potassium channel activity.

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EXAMPLE 9

Methods of Treatment

The present invention includes a treatment that inhibits oxidation associated diseases in a subject such as an animal, for example a rat or human. The method includes administering the antioxidant drugs of the present invention, or a combination of the antioxidant drug and one or more other pharmaceutical agents, to the subject in a pharmaceutically compatible carrier and in an effective amount to inhibit the development or progression of oxidation associated diseases. Although the treatment can be used prophylactically in any patient in a demographic group at significant risk for such diseases, subjects can also be selected using more specific criteria, such as a definitive diagnosis of the condition. The administration of any exogenous antioxidant cannabinoid would inhibit the progression of the oxidation associated disease as compared to a subject to whom the cannabinoid was not administered. The antioxidant effect, however, increases with the dose of the cannabinoid.

The vehicle in which the drug is delivered can include pharmaceutically acceptable compositions of the drugs of the present invention using methods well known to those with skill in the art. Any of the common carriers, such as sterile saline or glucose solution, can be utilized with the drugs provided by the invention. Routes of administration include but are not limited to oral, intracranial ventricular (icv), intrathecal (it), intravenous (iv), parenteral, rectal, topical ophthalmic, subconjunctival, nasal, aural, sub-lingual (under the tongue) and transdermal. The antioxidant drugs of the invention may be administered intravenously in any conventional medium for intravenous injection such as an aqueous saline medium, or in blood plasma medium. Such medium may also contain conventional pharmaceutical adjunct materials such as, for example, pharmaceutically acceptable salts to adjust the osmotic pressure, lipid carriers such as cyclodextrins, proteins such as serum albumin, hydrophilic agents such as methyl cellulose, detergents, buffers, preservatives and the like. Given the low solubility of many cannabinoids, they may be suspended in sesame oil.

Given the excellent absorption of the compounds of the present invention via an inhaled route, the compounds may also be administered as inhalants, for example in pharmaceutical aerosols utilizing solutions, suspensions, emulsions, powders and semisolid preparations of the type more fully described in *Remington: The Science and Practice of Pharmacy* (19th Edition, 1995) in chapter 95. A particular inhalant form is a metered dose inhalant containing the active ingredient, in a suspension or a dispersing agent (such as sorbitan trioleate, oleyl alcohol, oleic acid, or lecithin, and a propellant such as 12/11 or 12/114).

Embodiments of the invention comprising pharmaceutical compositions can be prepared with conventional pharmaceutically acceptable carriers, adjuvants and counterions as would be known to those of skill in the art. The compositions are preferably in the form of a unit dose in solid, semi-solid and liquid dosage forms such as tablets, pills, powders, liquid solutions or

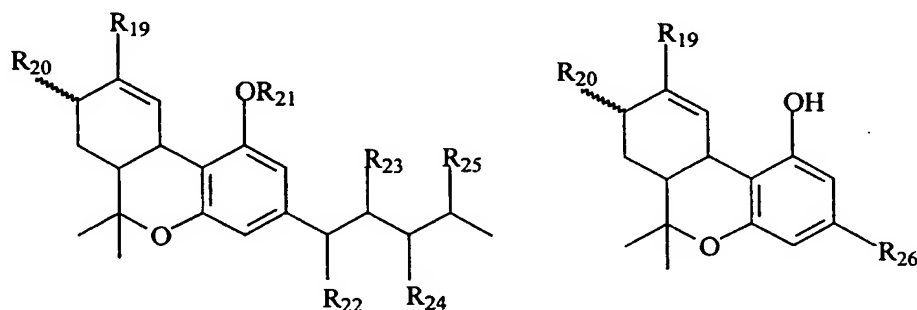
suspensions, injectable and infusible solutions, for example a unit dose vial, or a metered dose inhaler. Effective oral human dosage ranges for cannabidiol are contemplated to vary from about 1-40 mg/kg, for example 5-20 mg/kg, and in particular a dose of about 20 mg/kg of body weight.

If the antioxidant drugs are to be used in the prevention of cataracts, they may be administered in the form of eye drops formulated in a pharmaceutically inert, biologically acceptable carrier, such as isotonic saline or an ointment. Conventional preservatives, such as benzalkonium chloride, can also be added to the formulation. In ophthalmic ointments, the active ingredient is admixed with a suitable base, such as white petrolatum and mineral oil, along with antimicrobial preservatives. Specific methods of compounding these dosage forms, as well as appropriate pharmaceutical carriers, are known in the art. *Remington: The Science and Practice of Pharmacy*, 19th Ed., Mack Publishing Co. (1995), particularly Part 7.

The compounds of the present invention are ideally administered as soon as a diagnosis is made of an ischemic event, or other oxidative insult. For example, once a myocardial infarction has been confirmed by electrocardiograph, or an elevation in enzymes characteristic of cardiac injury (e.g. CKMB), a therapeutically effective amount of the cannabinoid drug is administered. A dose can also be given following symptoms characteristic of a stroke (motor or sensory abnormalities), or radiographic confirmation of a cerebral infarct in a distribution characteristic of a neurovascular thromboembolic event. The dose can be given by frequent bolus administration, or as a continuous IV dose. In the case of cannabidiol, for example, the drug could be given in a dose of 5 mg/kg active ingredient as a continuous intravenous infusion; or hourly intramuscular injections of that dose.

EXAMPLE 10

The following table lists examples of some dibenzopyran cannabinoids that may be useful as antioxidants in the method of the present invention.



- 25 -

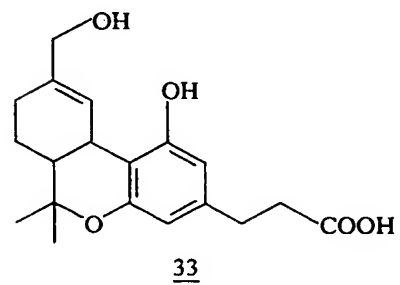
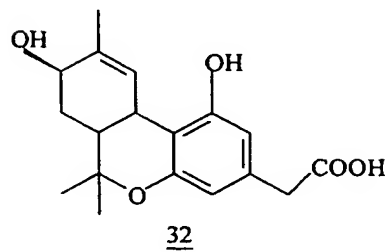
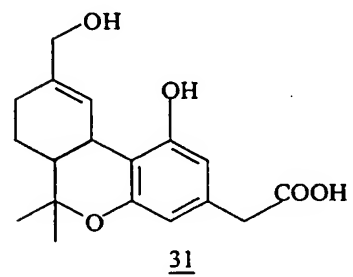
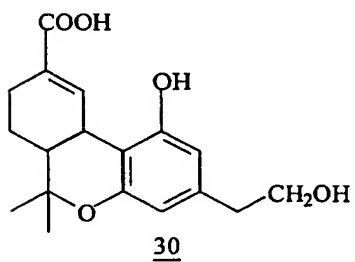
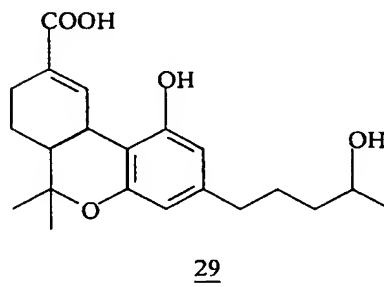
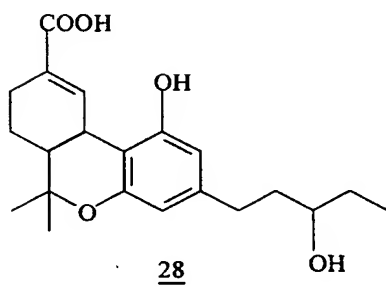
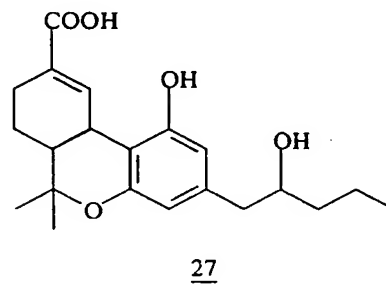
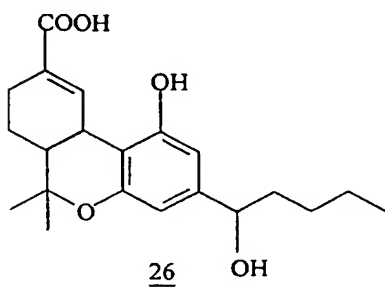
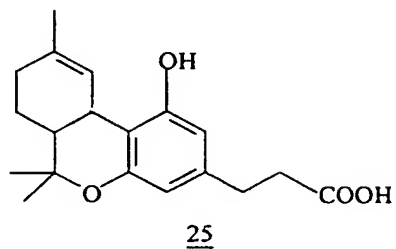
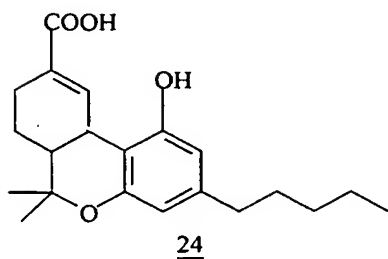
| Compound | | R ₁₉ | R ₂₀ | R ₂₁ | R ₂₂ | R ₂₃ | R ₂₄ | R ₂₅ | R ₂₆ |
|-------------|--|--------------------|-------------------|-----------------|-------------------|-----------------|-----------------|-----------------|------------------------------------|
| H <u>5</u> | 7-OH- Δ^1 -THC | CH ₂ OH | H | H | H | H | H | H | C ₅ H ₁₁ |
| H <u>6</u> | 6 α -OH- Δ^1 -THC | CH ₃ | α -OH | | | | | | |
| H <u>7</u> | 6 β -OH- Δ^1 -THC | CH ₃ | β -OH | | | | | | |
| H <u>8</u> | 1"-OH- Δ^1 -THC | CH ₃ | | | OH | | | | |
| H <u>9</u> | 2"-OH- Δ^1 -THC | CH ₃ | | | | OH | | | |
| <u>10</u> | 3"-OH- Δ^1 -THC | CH ₃ | | | | | OH | | |
| <u>11</u> | 4"-OH- Δ^1 -THC | CH ₃ | | | | | | OH | |
| H <u>12</u> | 6 α ,7-diOH- Δ^1 -THC | CH ₂ OH | α -OH | | | | | | |
| H <u>13</u> | 6 ν ,7-diOH- Δ^1 -THC | CH ₂ OH | β -OH | | | | | | |
| <u>14</u> | 1",7-diOH- Δ^1 -THC | CH ₂ OH | | | OH | | | | |
| H <u>15</u> | 2",7-diOH- Δ^1 -THC | CH ₂ OH | | | | OH | | | |
| H <u>16</u> | 3",7-diOH- Δ^1 -THC | CH ₂ OH | | | | | OH | | |
| H <u>17</u> | 4",7-diOH- Δ^1 -THC | CH ₂ OH | | | | | | OH | |
| <u>18</u> | 1",6 β -diOH- Δ^1 -THC | CH ₃ | β -OH | | OH | | | | |
| <u>19</u> | 1",3"-diOH- Δ^1 -THC | CH ₃ | | | OH | | OH | | |
| <u>20</u> | 1",6 α ,7-triOH- Δ^1 -THC | CH ₂ OH | α -OH | | OH | | | | |
| H <u>21</u> | Δ^1 -THC-6-one | CH ₃ | =O | | | | | | |
| <u>22</u> | Epoxyhexahydrocannabinol (EHHC)* | CH ₃ | | | | | | | |
| <u>23</u> | 7-oxo- Δ^1 -THC | CHO | | | | | | | |
| H <u>24</u> | Δ^1 -THC-7-oic acid | COOH | | | | | | | |
| H <u>25</u> | Δ^1 -THC-3"-oic acid | CH ₃ | | | | | | | C ₂ H ₄ COOH |
| H <u>26</u> | 1"-OH- Δ^1 -THC-7"-oic acid | COOH | | | OH | | | | |
| H <u>27</u> | 2"-OH- Δ^1 -THC-7"-oic acid | COOH | | | | OH | | | |
| H <u>28</u> | 3"-OH- Δ^1 -THC-7"-oic acid | COOH | | | | | OH | | |
| H <u>29</u> | 4"-OH- Δ^1 -THC-7"-oic acid | COOH | | | | | | OH | |
| H <u>30</u> | 3",4",5"-trisor-2"-OH- Δ^1 -THC-7-oic acid | COOH | | | | | | | C ₂ H ₄ OH |
| H <u>31</u> | 7-OH- Δ^1 -THC-2"-oic acid | CH ₂ OH | | | | | | | CH ₂ COOH |
| H <u>32</u> | 6 β -OH- Δ^1 -THC-2"-oic acid | CH ₃ | β -OH | | | | | | CH ₂ COOH |
| H <u>33</u> | 7-OH- Δ^1 -THC-3"-oic acid | CH ₂ OH | | | | | | | C ₂ H ₄ COOH |
| H <u>34</u> | 6 β -OH- Δ^1 -THC-3"-oic acid | CH ₃ | β -OH | | | | | | C ₂ H ₄ COOH |
| H <u>35</u> | 6 α -OH- Δ^1 -THC-4"-oic acid | CH ₃ | α -OH | | | | | | C ₃ H ₆ COOH |
| H <u>36</u> | 2",3"-dehydro-6 β -OH- Δ^1 -THC-4"-oic acid | CH ₃ | α -OH | | | | | | C ₃ H ₄ COOH |
| H <u>37</u> | Δ^1 -THC-1",7-dioic acid | COOH | | | | | | | COOH |
| H <u>38</u> | Δ^1 -THC-2",7-dioic acid | COOH | | | | | | | CH ₂ COOH |
| H <u>39</u> | Δ^1 -THC-3",7-dioic acid | COOH | | | | | | | C ₂ H ₄ COOH |
| H <u>40</u> | Δ^1 -THC-4",7-dioic acid | COOH | | | | | | | C ₃ H ₆ COOH |
| H <u>41</u> | 1",2"-dehydro- Δ^1 -THC-3",7-dioic acid | COOH | | | | | | | C ₂ H ₂ COOH |
| H <u>42</u> | Δ^1 -THC-glucuronic acid | CH ₃ | | | gluc [†] | | | | |
| H <u>43</u> | Δ^1 -THC-7-oic acid glucuronide | COO | gluc [†] | | | | | | |

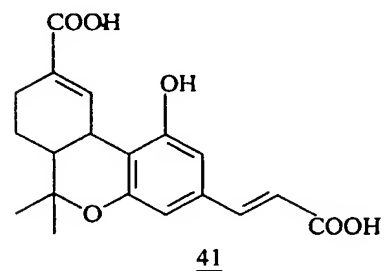
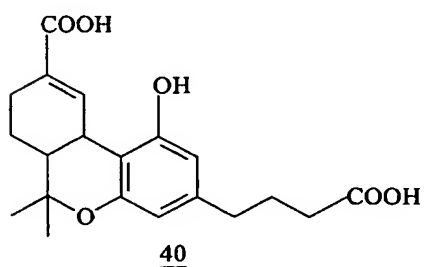
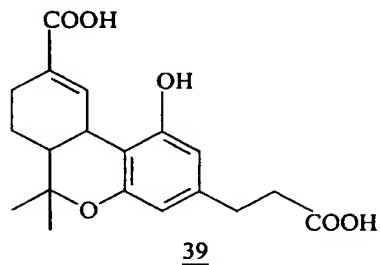
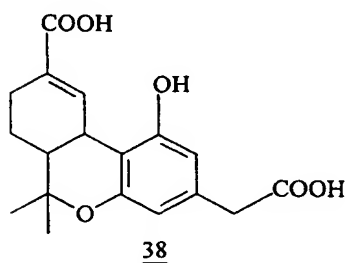
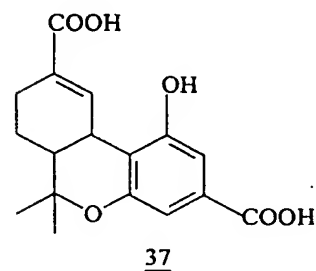
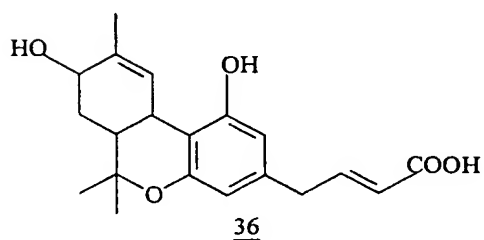
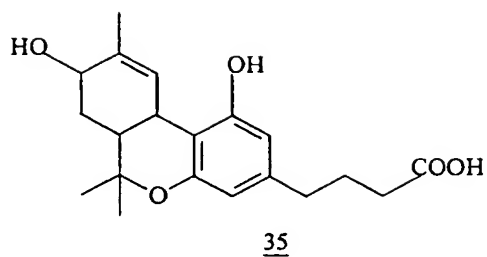
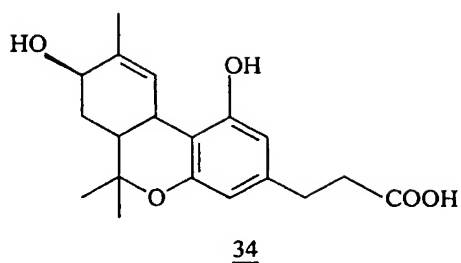
* Epoxy group in C-1 and C-2 positions

† Glucuronide

Note: R-group substituents are H if not indicated otherwise.

Chemical structures of some of the dibenzopyran cannabinoids are shown below.

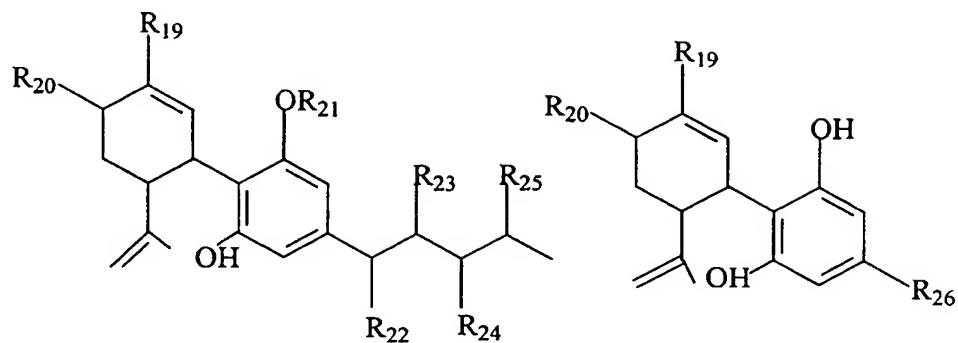




EXAMPLE 11

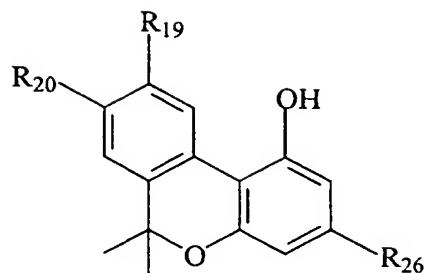
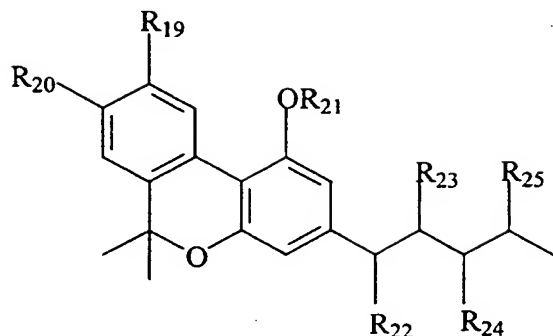
Examples of Structural Analogs of Cannabidiol

The following table lists examples of some cannabinoids which are structural
5 analogs of cannabidiol and that may be useful as antioxidants in the method of the present
invention. A particularly useful example is compound CBD, cannabidiol.



| Compound | R ₁₉ | R ₂₀ | R ₂₁ | R ₂₂ | R ₂₃ | R ₂₄ | R ₂₅ | R ₂₆ |
|---------------------------|--------------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|--|
| <u>44</u> CBD | CH ₃ | H | H | H | H | H | H | C ₅ H ₁₁ |
| <u>45</u> 7-OH-CBD | CH ₂ OH | | | | | | | |
| <u>46</u> 6 α - | CH ₃ | α -OH | | | | | | |
| <u>47</u> 6 β - | CH ₃ | β -OH | | | | | | |
| <u>48</u> 1"- | CH ₃ | | | OH | | | | |
| <u>49</u> 2"- | CH ₃ | | | | OH | | | |
| <u>50</u> 3"- | CH ₃ | | | | | OH | | |
| <u>51</u> 4"- | CH ₃ | | | | | | OH | |
| <u>52</u> 5"- | CH ₃ | | | | | | | C ₄ H ₈ CH ₂ OH |
| <u>53</u> 6,7-diOH-CBD | CH ₂ OH | OH | | | | | | |
| <u>54</u> 3",7-diOH-CBD | CH ₂ OH | | | | | OH | | |
| <u>55</u> 4",7-diOH-CBD | CH ₂ OH | | | | | | OH | |
| <u>56</u> CBD-7-oic acid | COOH | | | | | | | |
| <u>57</u> CBD-3"-oic acid | CH ₃ | | | | | | | C ₂ H ₄ COOH |

Note: R-group substituents are H if not indicated otherwise.



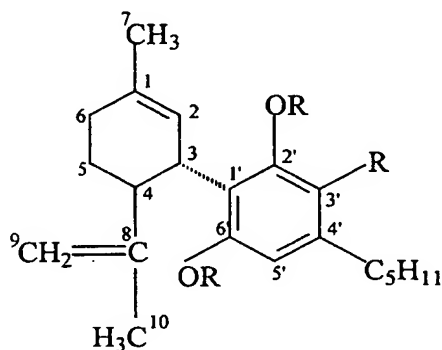
| Compound | R ₁₉ | R ₂₀ | R ₂₁ | R ₂₂ | R ₂₃ | R ₂₄ | R ₂₅ | R ₂₆ |
|---------------------------|--------------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|--|
| <u>58</u> CBN | CH ₃ | H | H | H | H | H | H | C ₅ H ₁₁ |
| <u>59</u> 7-OH-CBN | CH ₂ OH | | | | | | | |
| <u>60</u> 1"-OH-CBN | CH ₃ | | | OH | | | | |
| <u>61</u> 2"-OH-CBN | CH ₃ | | | | OH | | | |
| <u>62</u> 3"-OH-CBN | CH ₃ | | | | | OH | | |
| <u>63</u> 4"-OH-CBN | CH ₃ | | | | | | OH | |
| <u>64</u> 5"-OH-CBN | CH ₃ | | | | | | | C ₄ H ₉ CH ₂ OH |
| <u>65</u> 2"-7-diOH-CBN | CH ₂ OH | | | | OH | | | |
| <u>66</u> CBN-7-oic acid | COOH | | | | | | | |
| <u>67</u> CBN-1"-oic acid | CH ₃ | | | | | | | COOH |
| <u>68</u> CBN-3"-oic acid | CH ₃ | | | | | | | C ₂ H ₄ COOH |

Note: R-group substituents are H if not indicated otherwise.

The invention being thus described, variation in the materials and methods for practicing the invention will be apparent to one of ordinary skill in the art. Such variations are to be considered within the scope of the invention, which is set forth in the claims below.

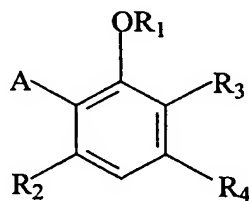
We claim:

1. A method of treating diseases caused by oxidative stress, comprising administering a therapeutically effective amount of a cannabinoid to a subject who has a disease
- 5 caused by oxidative stress.
2. The method of claim 1, wherein the cannabinoid is nonpsychoactive.
3. The method of claim 2, wherein the cannabinoid has a volume of distribution of 10 L/kg or more.
4. The method of claim 2, wherein the cannabinoid is not an antagonist at the
- 10 NMDA and AMPA receptors.
5. The method of claim 1, wherein the cannabinoid is



where R is selected from the group of H, substituted or unsubstituted alkyl, carboxyl, alkoxy, aryl, aryloxy, arylalkyl, halo or amino.

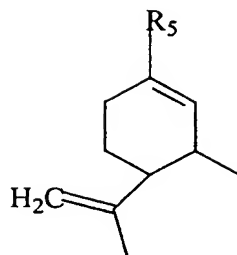
6. The method of claim 5, wherein R is selected from the group consisting of H,
- 15 and substituted or unsubstituted alkyl, or carboxyl, alkoxy.
7. The method of claim 2, wherein the cannabinoid is selected from the group



where

A is selected from the group of cyclohexyl and substituted or unsubstituted aryl,

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but not a pinene

R_1 is selected from the group of H, substituted or unsubstituted alkyl, and substituted or unsubstituted carboxyl;

5 R_2 is selected from the group of H, lower substituted or unsubstituted alkyl, and alkoxy;

R_3 is selected from the group of H, lower substituted or unsubstituted alkyl, and substituted or unsubstituted carboxyl;

10 R_4 is selected from the group of H, hydroxyl and lower substituted or unsubstituted alkyl; and

R_5 is selected from the group of H, hydroxyl and lower substituted or unsubstituted alkyl;

8. The method of claim 7, wherein

R_1 is selected from the group of lower alkyl, COOH and COCH₃;

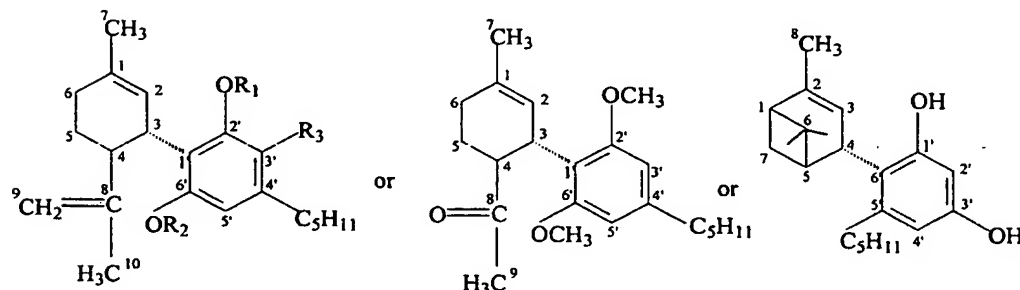
15 R_2 is selected from the group of unsubstituted C₁-C₅ alkyl, hydroxyl, methoxy and ethoxy;

R_3 is selected from the group of H, unsubstituted C₁-C₃ alkyl, and COCH₃;

R_4 is selected from the group of hydroxyl, pentyl, heptyl and diethylheptyl;

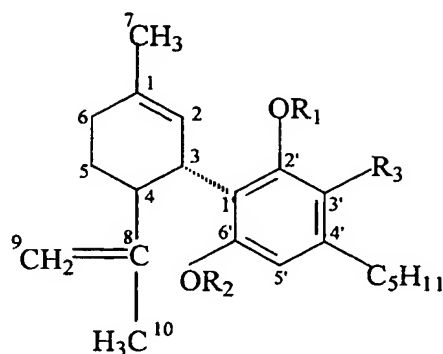
R_5 is selected from the group of hydroxyl and methyl;

20 9. The method of claim 1, wherein the cannabinoid is selected from the group of:



wherein R_1 , R_2 and R_3 are independently selected from the group consisting of H, CH₃, and COCH₃.

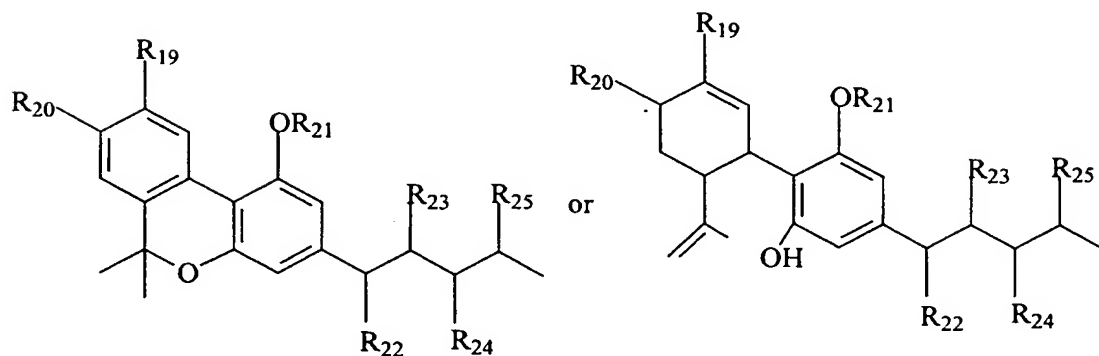
10. The method of claim 9, wherein the cannabinoid is selected from the group of:



where:

- a) $R_1 = R_2 = R_3 = H$
- b) $R_1 = R_3 = H, R_2 = CH_3$
- c) $R_1 = R_2 = CH_3, R_3 = H$
- d) $R_1 = R_2 = COCH_3, R_3 = H$
- e) $R_1 = H, R_2 = R_3 = COCH_3$

11. The method of claim 2, wherein the cannabinoid is selected from the group of:

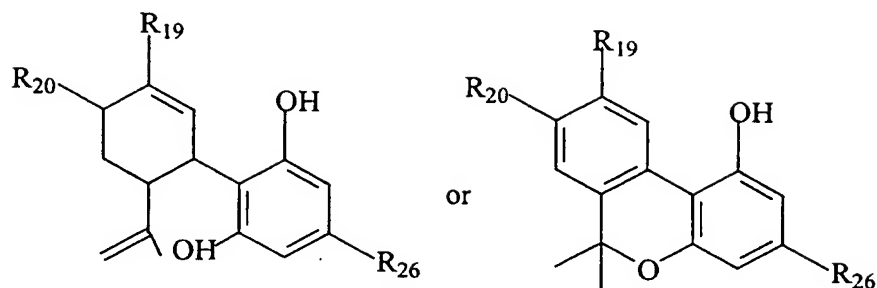


wherein R_{19} is H, lower alkyl, lower alcohol, or carboxyl; R_{20} is H or OH; and R_{21} - R_{25} are independently H or OH.

12. The method of claim 11, wherein R_{19} is H, CH_3 , CH_2OH , or $COOH$, and R_{20} - R_{25} are independently H or OH.

13. The method of claim 2, wherein the cannabinoid is

- 33 -



wherein R_{19} and R_{20} are H, and R_{26} is alkyl.

14. The method of claim 10, wherein the cannabinoid is cannabidiol.
15. A method of treating ischemic or neurodegenerative disease in the central nervous system of a subject, comprising administering to the subject a therapeutically effective amount of a cannabinoid of claim 5.
16. The method of claim 13, wherein the cannabinoid is not a psychoactive cannabinoid.
17. The method of claim 15, wherein the ischemic or neurodegenerative disease is selected from the group consisting of an ischemic infarct, Alzheimer's disease, Parkinson's disease, and human immunodeficiency virus dementia, Down's syndrome, and heart disease.
18. An assay for selecting a cannabinoid to use in treating a neurological disease, comprising determining whether the cannabinoid is an antioxidant.
19. A method of treating a neurological or ischemic disease, comprising using the assay of claim 18 to determine whether the cannabinoid is an antioxidant, and administering the cannabinoid if it is an antioxidant.
20. The method of claim 19, wherein the neurological or ischemic disease is a vascular ischemic event in the central nervous system or myocardium.
21. The method of claim 20, wherein the neurological disease is a thromboembolism in the brain.
22. A method of treating a disease with a cannabinoid, comprising determining whether the disease is caused by oxidative stress, and if the disease is caused by oxidative stress, administering the cannabinoid in a therapeutically effective antioxidant amount.
23. The method of claim 22, wherein the cannabinoid has a volume of distribution of at least 1.5 L/kg and substantially no activity at the cannabinoid receptor.
24. The method of claim 23, wherein the cannabinoid has a volume of distribution of at least 10 L/kg.
25. The method of claim 1, wherein the cannabinoid selectively inhibits an enzyme activity of 5- and 15-lipoxygenase more than an enzyme activity of 12-lipoxygenase.

- 34 -

26. An assay for selecting a cannabinoid to use in treating a neurological disease, comprising determining whether the cannabinoid selectively inhibits an enzyme activity of 5- and 15-lipoxygenase substantially more than the enzyme activity of an 12-lipoxygenase.

5 27. A method of treating a neurological or ischemic disease, comprising using the assay of claim 26 to determine whether the cannabinoid inhibits the enzyme activity of 5- and 15-lipoxygenase substantially more than 12-lipoxygenase, and administering the cannabinoid if it inhibits the enzyme activity of 5- and 15-lipoxygenase substantially more than 12-lipoxygenase.

28. The method of claim 27, wherein the neurological or ischemic disease is a vascular ischemic event in the central nervous system or myocardium.

10 29. The method of claim 28, wherein the neurological disease is a thromboembolism in the brain.

30. The method of claim 27, wherein the cannabinoid is administered during and after an ischemic injury.

15 31. The assay of claim 18 further comprising determining whether the cannabinoid selectively inhibits an enzyme activity of 5- and 15-lipoxygenase substantially more than the enzyme activity of an 12-lipoxygenase, and determining whether the cannabinoid has low NMDA antagonist activity.

32. The method of claim 31, further comprising determining whether the cannabinoid possess potent cannabinoid receptor activity.

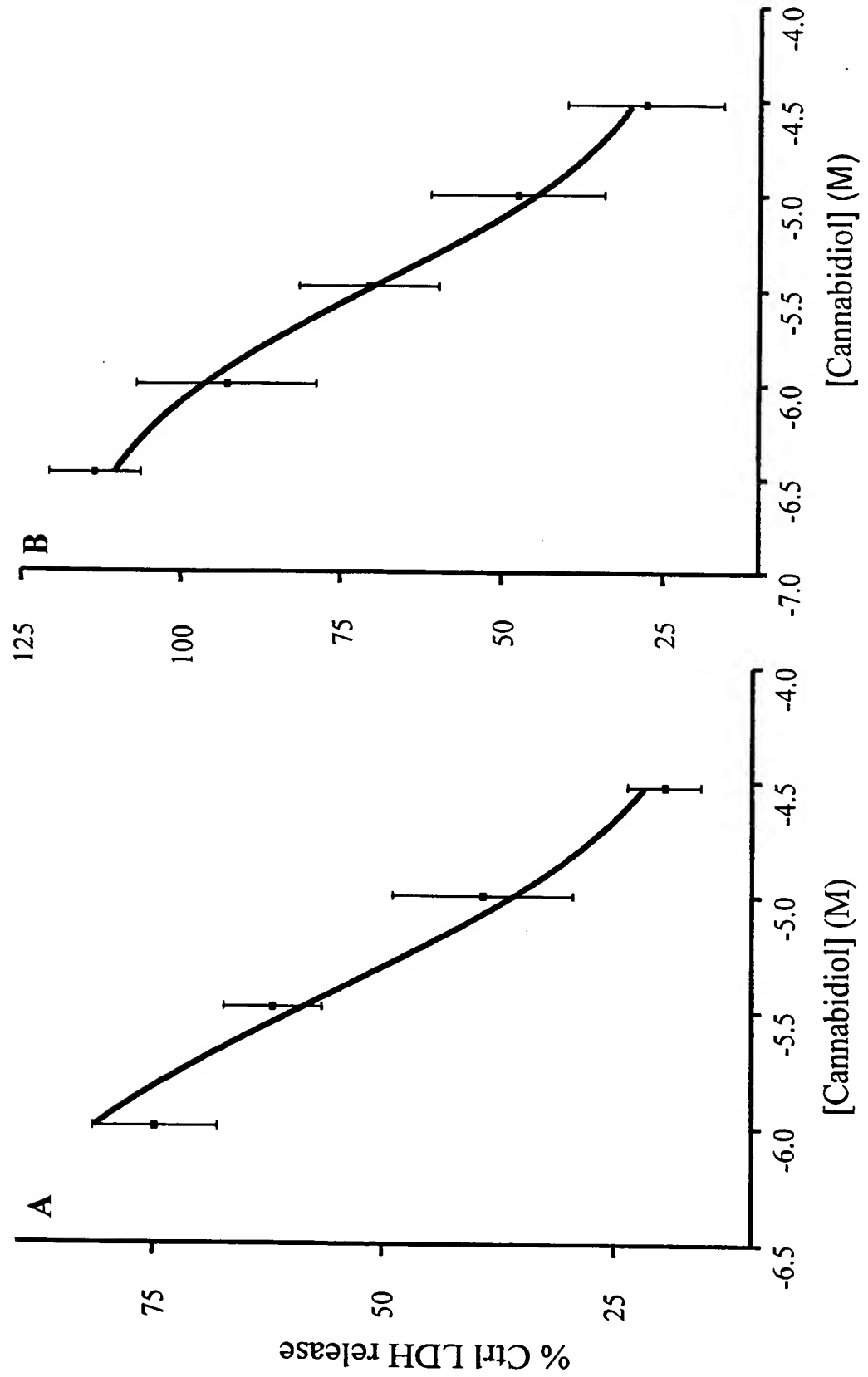
FIG. 1

FIG. 2

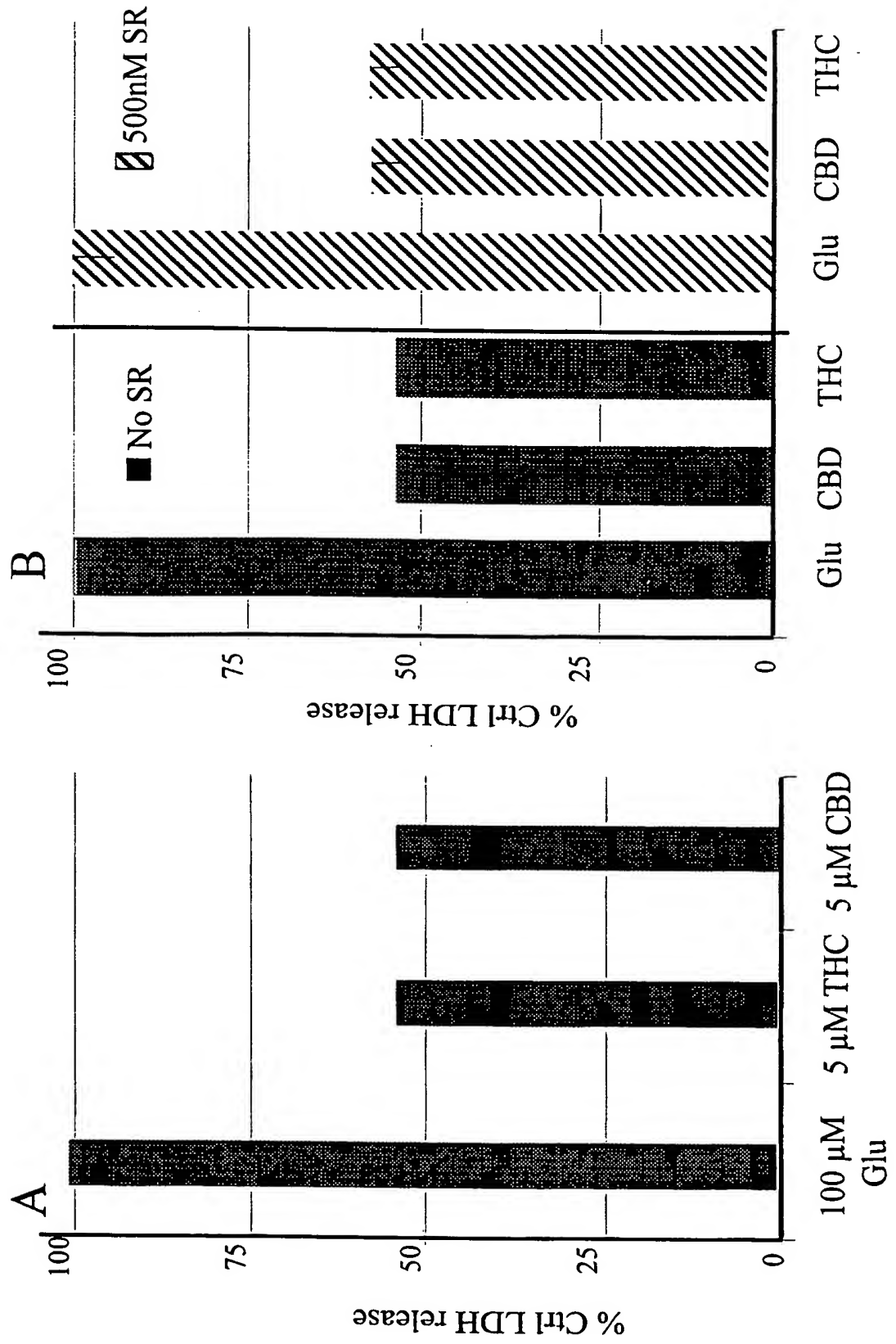


FIG. 3

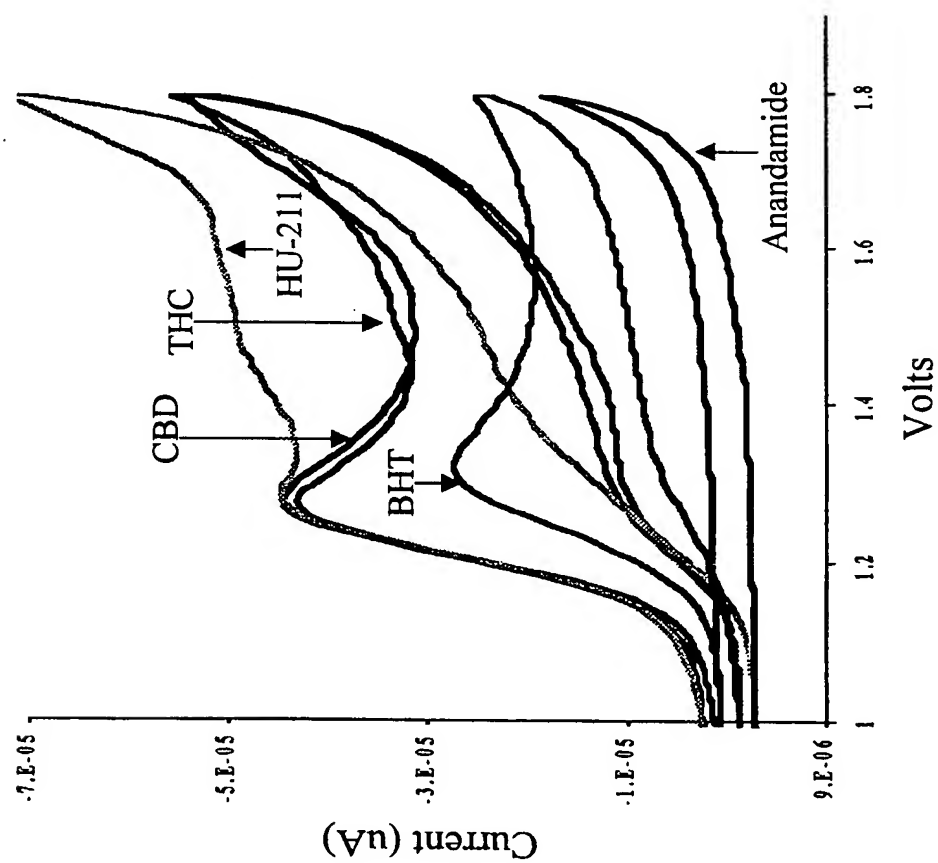


FIG. 4

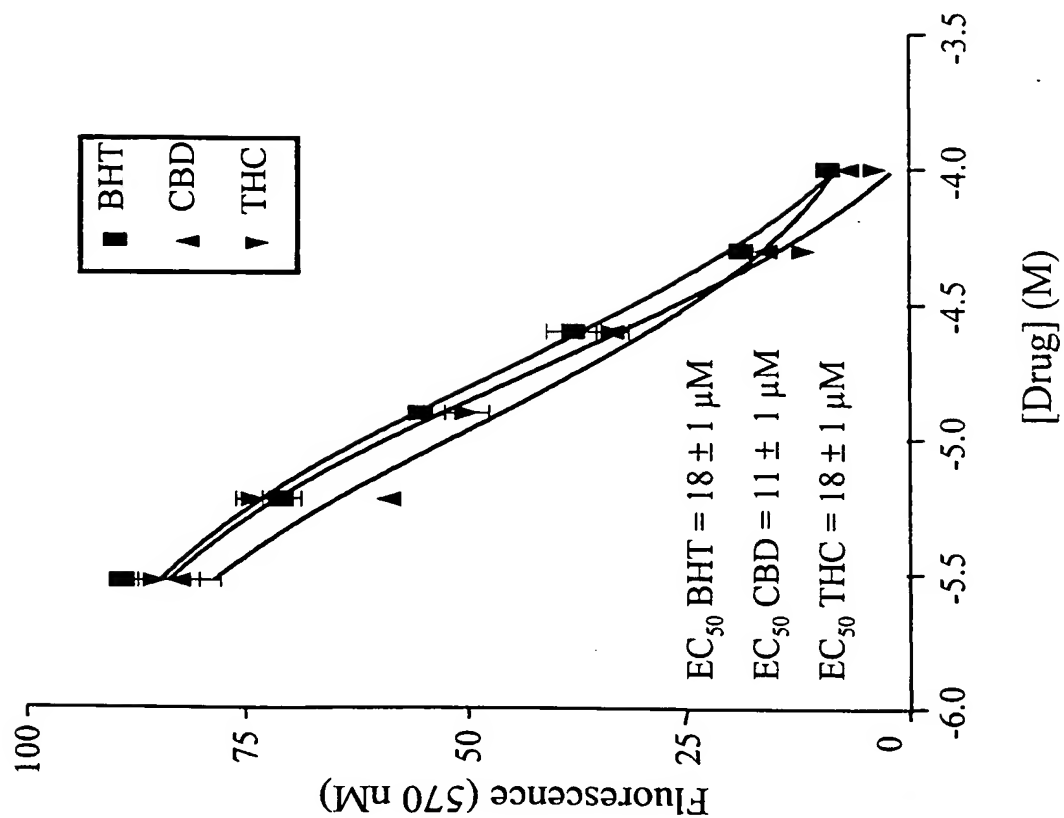


FIG. 5

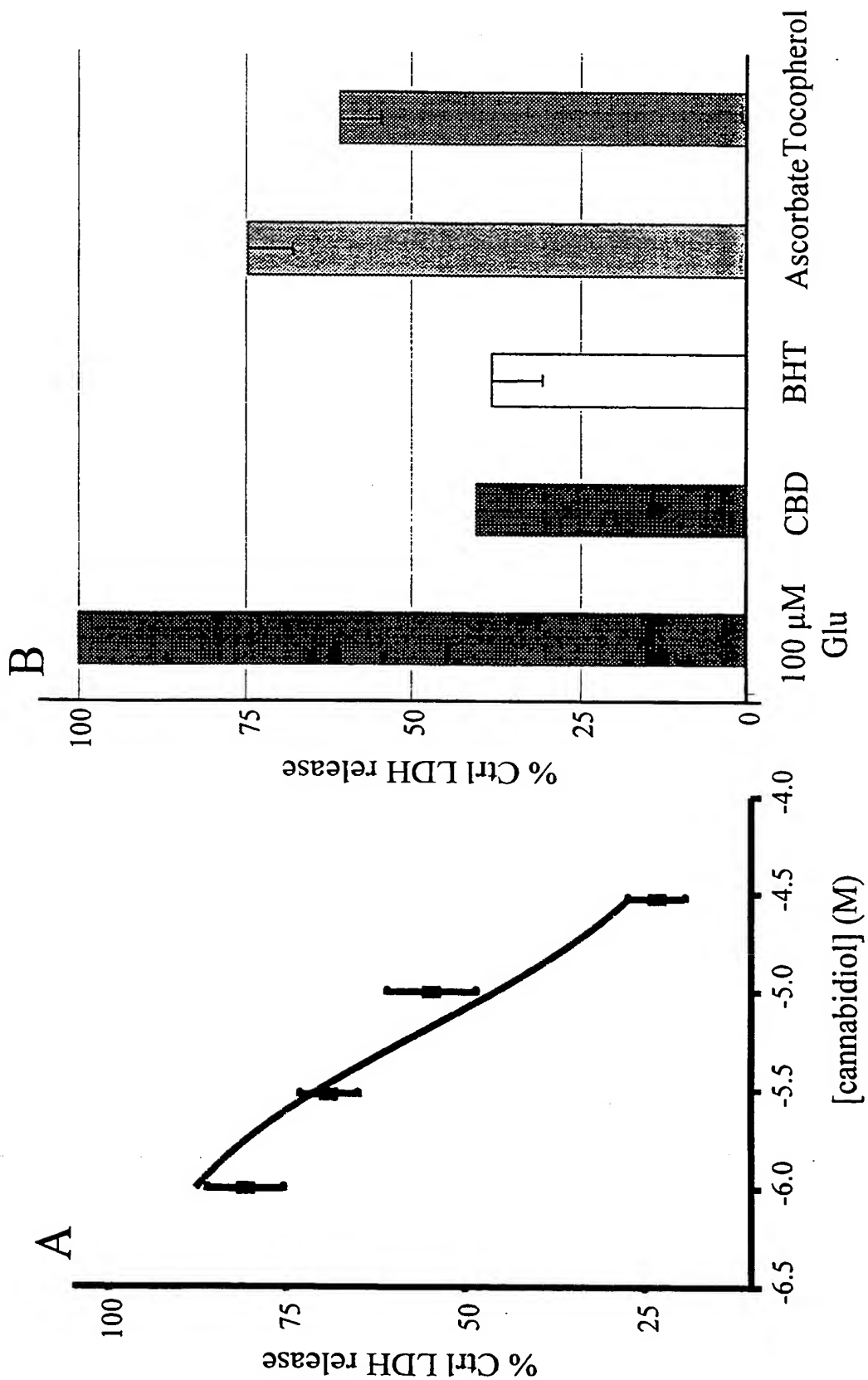


FIG. 6

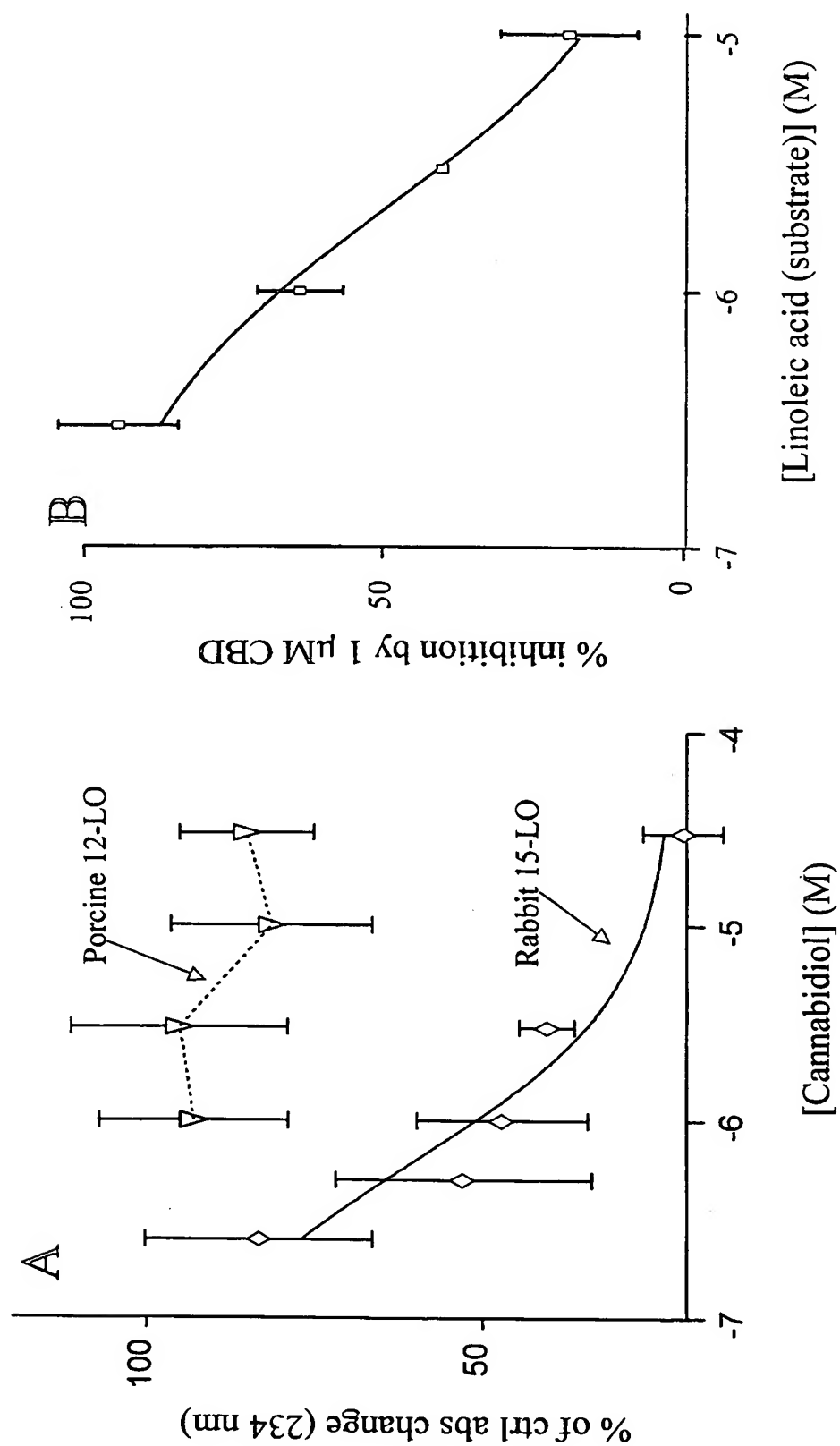


FIG. 7

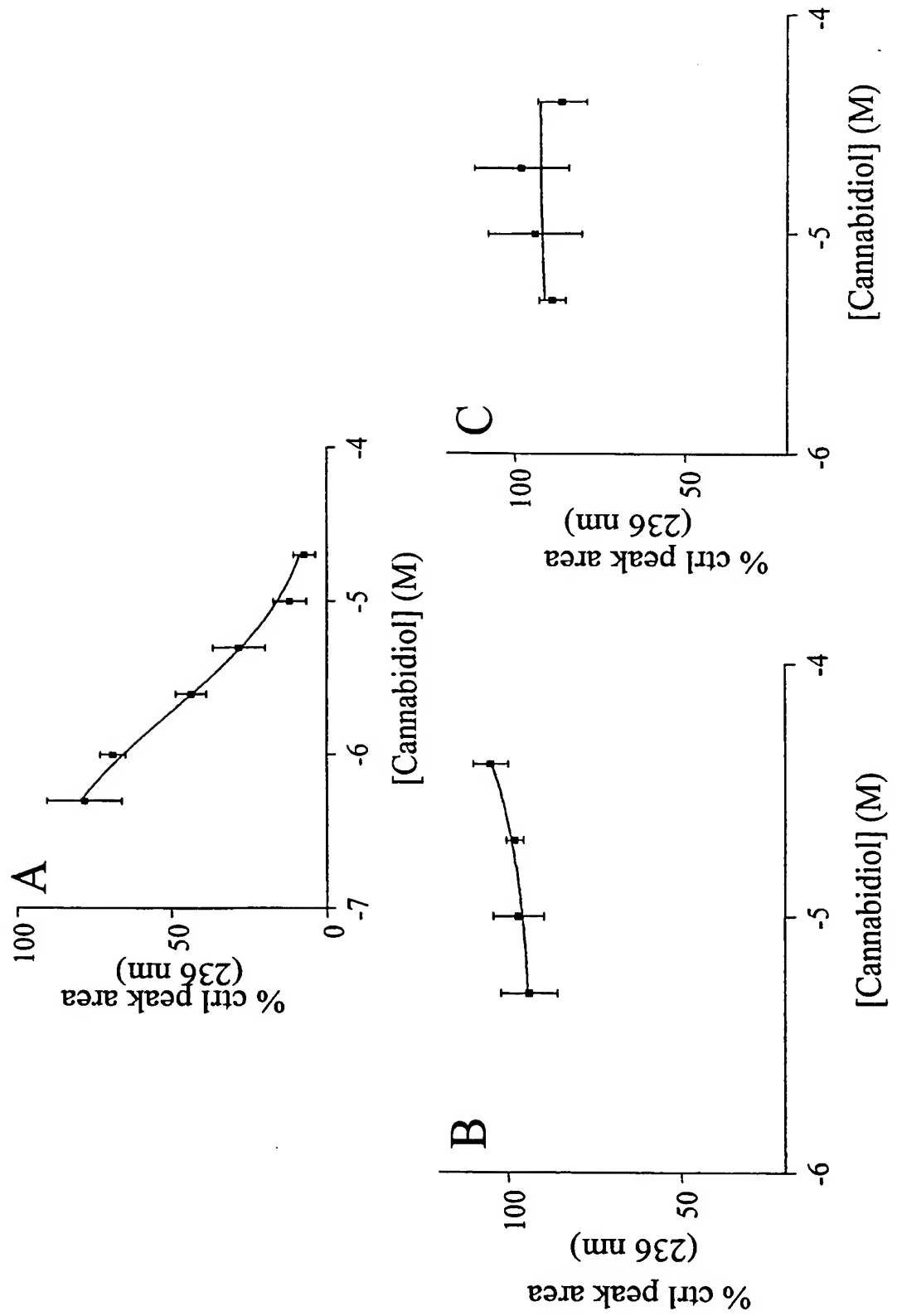
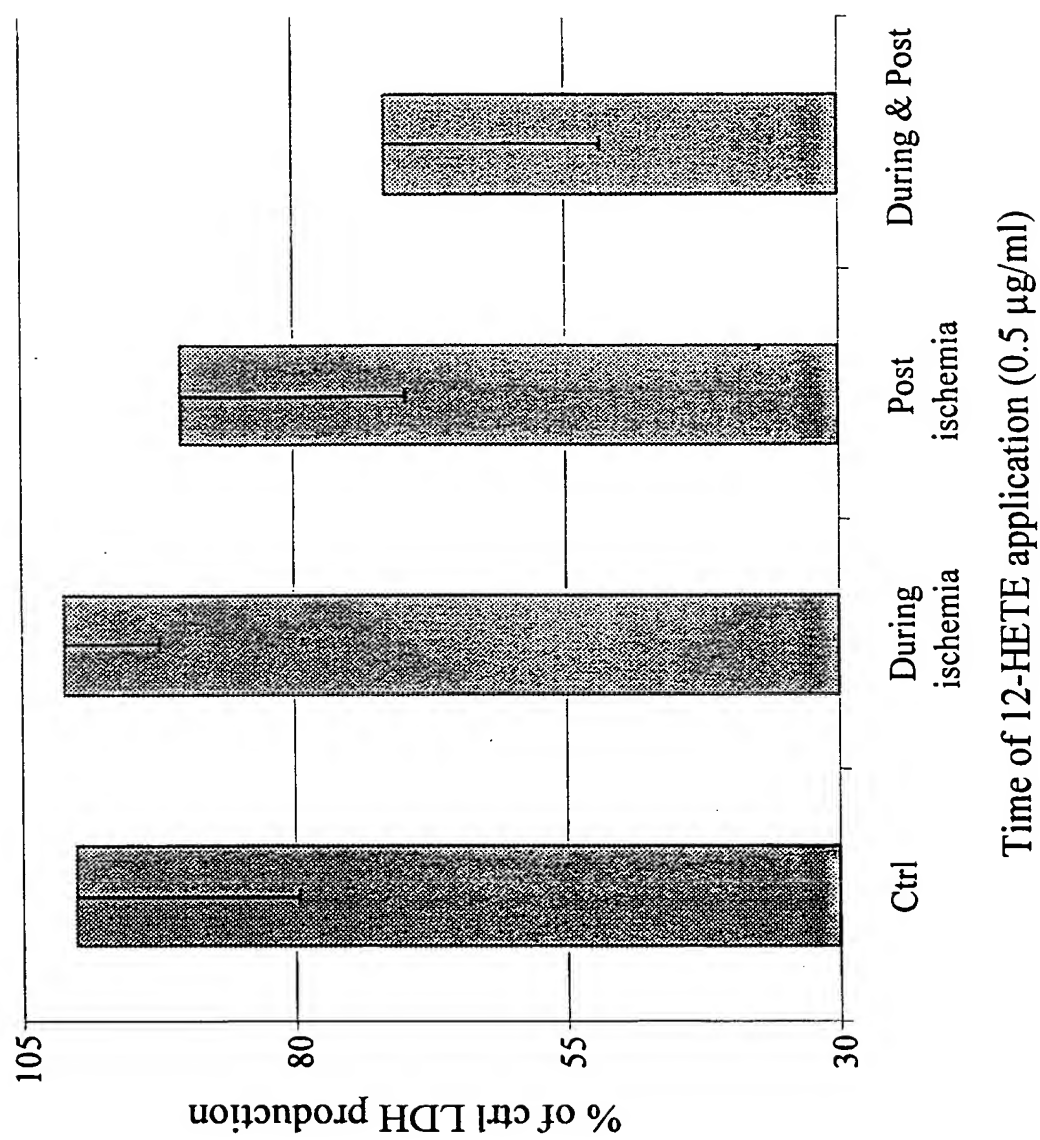


FIG. 8

INTERNATIONAL SEARCH REPORT

In International Application No
PCT/US 99/08769

A. CLASSIFICATION OF SUBJECT MATTER
IPC 6 A61K31/35 A61K31/05 A61K31/09 A61K31/12

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 6 A61K C07D

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

| Category | Citation of document, with indication, where appropriate, of the relevant passages | Relevant to claim No. |
|----------|---|-----------------------|
| P, X | HAMPSON ET AL.: "Cannabidiol and Tetrahydrocannabinol are neuroprotective antioxidants" PROC. NATL. ACAD. SCI. USA, vol. 95, July 1998 (1998-07), pages 8268-8273, XP002109751 the whole document | 1-32 |
| X | US 5 521 215 A (KLOOG YOEL ET AL) 28 May 1996 (1996-05-28) cited in the application column 3, line 37-45 column 4, line 25-29 column 6, line 21-29; claims; figures 1-13; examples | 1-3 |

☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

Special categories of cited documents:

"A" document defining the general state of the art which is not considered to be of particular relevance

"E" earlier document but published on or after the international filing date

"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

"&" document member of the same patent family

Date of the actual completion of the international search

20 July 1999

Date of mailing of the international search report

04/08/1999

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Authorized officer

Veronese, A

INTERNATIONAL SEARCH REPORT

International Application No.

PCT/US 99/08769

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

| Category | Citation of document, with indication, where appropriate, of the relevant passages | Relevant to claim No. |
|----------|--|-----------------------|
| X | ESHAR, N. S. ET AL.: "Neuroprotective and Antioxidant Activities of HU-211, a Novel NMDA Receptor Antagonist" EUR. J. PHARMACOL., vol. 283, no. 1-3, 1995, pages 19-29, XP002109752 | 1-3, 18-24 |
| Y | the whole document | 1-32 |
| X | ESTHER SHOHAMI ET AL.: "Oxidative Stress in Closed-Head Injury: Brain Antioxidant Capacity as an Indicator of Functional Outcome." JOURNAL OF CEREBRAL BLOOD FLOW AND METABOLISM, vol. 17, no. 10, October 1997 (1997-10), pages 1007-10019, XP002109753 | 1-3, 18-24 |
| Y | the whole document | 1-32 |
| X | SKAPER S. D. ET AL.: "The ALIAmide Palmitoylethanolamide and Cannabinoids, but not Anandamide, are Protective in a Delayed Postglutamate Paradigm of Excitotoxic Death in Cerebellar Neurons" PROC. NATL. ACAD. SCI. USA, vol. 93, 1997, pages 3984-3989, XP002109754 | 1-3 |
| Y | page 3896, column 1 table 1 | 1,32 |
| X | US 5 434 295 A (MECHOULAM RAPHAEL ET AL) 18 July 1995 (1995-07-18) cited in the application column 5, line 30-59: claims: examples | 7,8 |
| A | US 5 538 993 A (MECHOULAM RAPHAEL ET AL) 23 July 1996 (1996-07-23) cited in the application * See fig. 1A, compound 1,a * the whole document column 2, line 7,8 | 1-32 |

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US 99/08769

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☒ Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:
Remark: Although claim(s) 1-32
is(are) directed to a method of treatment of the human/animal
body, the search has been carried out and based on the alleged
effects of the compound/composition.
2. ☒ Claims Nos.: 7-8
because they relate to parts of the International Application that do not comply with the prescribed requirements to such
an extent that no meaningful International Search can be carried out, specifically:
see FURTHER INFORMATION sheet PCT/ISA/210
3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

1. ☐ As all required additional search fees were timely paid by the applicant, this International Search Report covers all
searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment
of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this International Search Report
covers only those claims for which fees were paid, specifically claims Nos.:
4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is
restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest.
- ☐ No protest accompanied the payment of additional search fees.

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

Continuation of Box I.2

Claims Nos.: 7-8

Present claims 7-8 relate to an extremely large number of possible compounds. Support within the meaning of Article 6 PCT and/or disclosure within the meaning of Article 5 PCT is to be found, however, for only a very small proportion of the compounds claimed. In the present case, the claims so lack support, and the application so lacks disclosure, that a meaningful search over the whole of the claimed scope is impossible. Consequently, the search has been carried out for those parts of the claims which appear to be supported and disclosed, namely those parts relating to the compounds in the examples and claims of the application

The applicant's attention is drawn to the fact that claims relating to inventions in respect of which no international search report has been established need not be the subject of an international preliminary examination (Rule 66.1(e) PCT). The applicant is advised that the EPO policy when acting as an International Preliminary Examining Authority is normally not to carry out a preliminary examination on matter which has not been searched. This is the case irrespective of whether or not the claims are amended following receipt of the search report or during any Chapter II procedure.

INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PCT/US 99/08769

| Patent document cited in search report | Publication date | Patent family member(s) | Publication date |
|---|---------------------|----------------------------|---------------------|
| US 5521215 A | 28-05-1996 | IL 92238 A | 31-07-1995 |
| | | US 5284867 A | 08-02-1994 |
| | | AU 690221 B | 23-04-1998 |
| | | AU 1743395 A | 21-08-1995 |
| | | CA 2183466 A | 10-08-1995 |
| | | EP 0765160 A | 02-04-1997 |
| | | JP 9511493 T | 18-11-1997 |
| | | SG 49625 A | 15-06-1998 |
| | | WO 9520958 A | 10-08-1995 |
| | | AT 119898 T | 15-04-1995 |
| | | AU 631262 B | 19-11-1992 |
| | | AU 6583490 A | 30-05-1991 |
| | | CA 2029419 A | 08-05-1991 |
| | | DE 69017839 D | 20-04-1995 |
| | | DE 69017839 T | 31-08-1995 |
| | | DK 427518 T | 24-07-1995 |
| | | EP 0427518 A | 15-05-1991 |
| | | ES 2071786 T | 01-07-1995 |
| | | JP 2038059 C | 28-03-1996 |
| | | JP 3209377 A | 12-09-1991 |
| | | JP 7068235 B | 26-07-1995 |
| | | KR 9505914 B | 07-06-1994 |
| US 5434295 A | 18-07-1995 | NONE | |
| US 5538993 A | 23-07-1996 | IL 99468 A | 30-09-1997 |
| | | US 5635530 A | 03-06-1997 |
| | | AU 664204 B | 09-11-1995 |
| | | AU 2678792 A | 05-04-1993 |
| | | CA 2118929 A | 18-03-1993 |
| | | EP 0642504 A | 15-03-1995 |
| | | JP 7505119 T | 08-06-1995 |
| | | WO 9305031 A | 18-03-1997 |